

HIV Diagnostic Testing

This is a PDF version of the following document:

Module 1: [Screening and Diagnosis](#)

Lesson 3: [HIV Diagnostic Testing](#)

You can always find the most up-to-date version of this document at

<https://www.hiv.uw.edu/go/screening-diagnosis/diagnostic-testing/core-concept/all>.

Background

HIV Testing and the Care Continuum

Diagnostic testing is the crucial first step in the HIV care continuum ([Figure 1](#)).^[1] Establishing a diagnosis of HIV has important implications for both HIV treatment and prevention. Accumulating evidence shows that persons with HIV who take antiretroviral therapy without interruption maintain suppressed plasma HIV RNA levels, do not transmit HIV to others sexually, and have better clinical outcomes.^[2,3,4,5,6] Improving rates of HIV testing and awareness of HIV status is critical because a high proportion of HIV transmissions occur from persons unaware of their HIV diagnosis.^[7] The highest HIV transmission rates occur among persons with acute HIV who are unaware of their diagnosis.^[7,8,9,10,11] The CDC estimated that in 2022, approximately 13% of people with HIV in the United States were unaware of their HIV diagnosis.^[12] Universal testing is also important because individuals who test negative but have a risk of acquiring HIV can be offered HIV risk-reduction counseling and preventative measures, including HIV preexposure prophylaxis (PrEP). Ideally, HIV testing, prevention, and treatment services are offered in the same setting in a “status-neutral” care model.^[13]

Approach to HIV Testing in the United States

In 2014, the CDC and the Association of Public Health Laboratories (APHL) released an HIV diagnostic algorithm to allow for more accurate diagnosis of acute HIV-1, improved ability to detect HIV type 2 (HIV-2), fewer indeterminate results, and faster turnaround time for completion of the testing algorithm.^[14] This diagnostic algorithm, which was updated in 2018, is discussed in detail below in the section CDC HIV Testing Algorithm. The HIV testing approach recommended by the CDC consists of initial screening with an HIV-1/2 antigen-antibody test, with follow-up testing of reactive samples using an HIV-1/HIV-2 differentiation antibody assay. The latter test can differentiate HIV-1 from HIV-2 and can provide antibody confirmation.^[14,15] Indeterminate or ambiguous results based on the initial HIV-1/2 antigen-antibody test and HIV-1/HIV-2 differentiation assay require further evaluation with an HIV nucleic acid test (NAT), such as an HIV-1 RNA PCR assay (or rarely an HIV-2 PCR).^[14] For more information about HIV-2, see the [HIV-2 Infection](#) lesson in this Key Populations module. From a practical standpoint, the same patient's blood sample can be used for the initial screening test and the HIV-1/HIV-2 differentiation assay. When using point-of-care sampling, such as an oral swab or fingerstick blood sample, the confirmatory testing requires obtaining an additional sample.

Clinical Laboratory Improvement Amendments (CLIA) Criteria

With a range of HIV diagnostic tests now available, the testing process can occur in a wide range of clinical and nonclinical settings. Most HIV testing is performed in a laboratory, and the time required to perform the

testing varies significantly, but some laboratory tests can be performed in less than an hour. Several point-of-care, single-use, rapid tests are now available that can be performed in clinical or nonclinical settings. In the United States, the Centers for Medicare and Medicaid Services (CMS) regulates all clinical laboratory testing through the Clinical Laboratory Improvement Amendments (CLIA). As part of this process, CLIA has established three levels of test complexity categories and this applies to the different HIV testing systems:

- **Waived:** These tests are considered simple to perform, with a low risk of an incorrect result, and can be performed with minimal training; specimens do not require centrifugation for testing.
- **Moderate Complexity:** Although these tests are considered simple to perform, the testing involves using plasma or serum specimens, and program participation in an external proficiency testing program.
- **High Complexity:** These tests involve multiple-step protocols and require trained laboratory personnel to perform, participation in an external proficiency testing program, and frequent checks on quality control.

Timing of Laboratory Markers following HIV Infection

Fiebig Staging System

Laboratory markers of HIV appear in a consistent sequence after infection occurs and delineate the period from initial exposure to established HIV. The Fiebig staging system, first published in 2003, defines 6 distinct stages of initial HIV infection that follow the eclipse phase, and these stages range from the emergence of HIV RNA (Stage I) to full Western blot reactivity (Stage VI) ([Figure 2](#)).^[16]

Early HIV Test Reactivity and Terminology

The CDC/APHL document and the Adult and Adolescent ART Guidelines have defined the following laboratory-based and clinically relevant terms related to acute HIV.^[14,17] Note that some of these terms are not standardized, and alternative terminology, such as primary HIV (instead of acute HIV), is often used.

- **Eclipse Phase:** The eclipse phase is the initial interval after HIV infection when no existing diagnostic test, including the HIV NAT, is capable of detecting HIV ([Figure 3](#)). During this phase, even the HIV RNA PCR assays, which are the first tests that can detect HIV following HIV acquisition, are negative because the HIV RNA levels have not yet reached levels detectable by standard laboratory assays.
- **Window Period:** The HIV seroconversion window period refers to the interval between HIV acquisition and the first detection of anti-HIV antibodies; the length of the window period varies depending on the specific HIV antibody assay used ([Figure 4](#)).
- **Acute HIV:** The term acute HIV (also referred to as primary HIV) typically describes the interval between the detection of HIV RNA and the detection of anti-HIV antibodies. During acute HIV infection, the HIV RNA is always detectable, and the HIV p24 antigen is often positive. People with acute HIV may have an abrupt onset of clinical symptoms.
- **Recent Infection:** The term recent infection usually describes the period after acute HIV when anti-HIV antibodies are developing out to 6 months after HIV acquisition.
- **Early Infection:** Early infection is generally used to describe both acute and recent HIV time periods, which extend out to 6 months after HIV acquisition.
- **Established HIV Infection:** The term established infection refers to the time after early HIV, during which anti-HIV IgG antibody responses have fully developed.

Timing of HIV Test Reactivity with Modern Assays

The CDC and Association of Public Health Laboratories (APHL) document, based on several CDC-related studies, have outlined the sequence of contemporary laboratory markers that turn positive following the acquisition of HIV ([Figure 5](#)).^[18,19] Following HIV acquisition, HIV RNA becomes detectable on standard laboratory tests approximately 10 days after infection.^[16] The next marker to appear is p24 antigen, which typically reaches detectable levels about 7 days after the emergence of HIV-1 RNA, with a positive p24 antigen test usually developing when the HIV-1 RNA exceeds 20,000-30,000 copies/mL.^[14,17] Next, the EIA (IgM antibody test) turns positive approximately 5 days later, and the IgM antibodies are gradually replaced by IgG antibodies, which appear 2 to 6 weeks after initial HIV RNA detection.^[16]

Tests Used for the Diagnosis of HIV

Updated Nomenclature for HIV Serologic Tests

In the United States, the Food and Drug Administration (FDA)-approved HIV serologic tests have historically been categorized as first-, second-, third-, and fourth-generation tests, based on evolving techniques and significant improvement in assay sensitivity.[20,21] With many of the new and improved HIV assays, a clear distinction between the HIV generations has blurred, and thus, use of the HIV test “generation” nomenclature is no longer recommended.[22] The first- and second-generation antibody assays are now referred to as IgG-sensitive tests, third-generation assays as IgM/IgG-sensitive tests, and fourth-generation as antigen-antibody immunoassays (Figure 6).[14,20,21,23] Further, the use of the term “rapid HIV test” to describe point-of-care tests is no longer recommended, since a number of instrumented, laboratory-based tests now have the capacity to generate HIV test results in less than one hour. Instead, the type of test is considered either laboratory-based or point-of-care.[21,22]

Summary of Test Methods used for Diagnosing HIV

The major tests used as screening tests for diagnosing HIV are (1) HIV antigen-antibody laboratory-based tests, (2) HIV antigen-antibody point-of-care tests, (3) HIV antibody laboratory-based tests, and (4) HIV antibody point-of-care tests.[24,25] There is also one FDA-approved HIV antibody test for use and interpretation at home.[26] Further, HIV diagnostic testing includes HIV-1/2 differentiation assays and HIV nucleic acid tests (NATs). The NAT testing includes options for qualitative HIV RNA or proviral DNA PCR assays or a quantitative RNA PCR assay, which is commonly referred to as a viral load test. The following summarizes the major FDA-approved HIV screening and differentiation assays.[15,21,25]

HIV Antigen-Antibody Laboratory-Based Tests

The HIV antigen-antibody laboratory-based immunoassays are the preferred screening tests for HIV. These immunoassays detect HIV-1 p24 (capsid) antigen and antibodies (IgM and IgG) to HIV-1 and HIV-2 (Figure 7).[14,21,27] The HIV-1/2 antigen-antibody immunoassays detect HIV significantly earlier than laboratory-based antibody tests, point-of-care antigen-antibody tests, and point-of-care HIV antibody tests.[21,28] All reactive HIV-1/2 antigen-antibody tests require confirmatory testing. None of the HIV-1/2 antigen-antibody immunoassays can detect HIV-2 core antigen (p26 antigen), but cross-reactivity to HIV-1 p24 antigen can occur in persons with HIV-2.

The following list (in alphabetical order) summarizes the laboratory-based HIV-1/2 antigen-antibody immunoassays that are FDA-approved for use in the United States:[24]

- **ADVIA Centaur HIV Ag/Ab Combo (CHIV) Assay:** This two-wash antigen-antibody sandwich immunoassay detects HIV-1 p24 antigen, antibodies to HIV-1, and antibodies to HIV-2.[29] This assay does not differentiate between p24 antigen and HIV antibodies. The ADVIA Centaur HIV Ag/Ab Combo Assay was evaluated in more than 7,000 samples and found to have a sensitivity of 98.4% and specificity of 99.7%.[30] This test is a chemiluminescent microparticle immunoassay (CMIA), and it requires less than 1 hour to perform.
- **ARCHITECT HIV Ag/Ab Combo:** This laboratory assay can detect HIV-1 p24 antigen, antibodies to HIV-1 (groups O and M), and antibodies to HIV-2.[31] This test does not distinguish antigen from antibody, nor does it distinguish HIV-1 from HIV-2. The ARCHITECT HIV Ag/Ab Combo assay has been evaluated in several studies and has demonstrated detection rates over 99% for established HIV infection and detection rates from 80% to 96% for acute HIV (with specificity above 98% in both established and acute HIV infection).[32,33,34] The ARCHITECT HIV Ag/Ab Combo is a CMIA, and it takes less than 30 minutes to perform.
- **BioPlex 2200 HIV Ag-Ab Assay:** This laboratory assay detects HIV-1 p24 antigen, antibodies to HIV-1 (groups O and M), and antibodies to HIV-2.[35] An advantage of this test is it can differentiate

between HIV-1 p24 antigen and antibodies to HIV-1 or HIV-2 and thus can help identify persons recently infected with HIV-1. Most other laboratory-based HIV antigen-antibody tests do not distinguish reactivity to p24 antigen from reactivity to HIV antibodies. Investigators prospectively tested the BioPlex 2200 HIV Ag-Ab Assay on 1,505 routine serum samples, and the assay had a sensitivity of 100% and specificity of 99.5%.[\[36\]](#) Additional testing on samples known to be positive for HIV-1, HIV-2, or both showed the differentiation capability of the assay was 100% for HIV-1, 90.7% for HIV-2, 100% for both HIV-1 and HIV-2, and 90.9% for early HIV infection.[\[36\]](#) This test is a multiplex flow immunoassay, and it takes 45 minutes to perform. A disadvantage of this test is it requires specialized equipment and specially trained technicians.

- **Elecsys HIV Combi PT:** This laboratory assay can detect HIV-1 p24 antigen, antibodies to HIV-1 (groups O and M), and antibodies to HIV-2.[\[37\]](#) A reactive test does not distinguish p24 antigen from antibodies to HIV, and it does not distinguish HIV-1 from HIV-2. This test is an electrochemiluminescence immunoassay (ECLIA), and it takes 27 minutes to perform.
- **Elecsys HIV Duo:** The laboratory-based ECLIA test is a double-sandwich immunoassay that uses monoclonal antibodies to detect p24 antigen and recombinant antigens to detect antibodies to HIV.[\[38\]](#) This assay evaluates serum or plasma samples for p24 antigen and antibodies to HIV-1 and HIV-2.[\[38\]](#) An advantage of this assay is that it differentiates p24 antigen reactivity from HIV-1 or HIV-2 antibody reactivity (the test gives an overall reactive or nonreactive result plus a sub-result that shows whether the p24 antigen or anti-HIV antibody, & or both, were positive). Although this test is one of a few antigen-antibody tests that distinguish between p24 antigen and anti-HIV antibody reactivity, it does not distinguish positive HIV-1 antibodies from positive HIV-2 antibodies.[\[38\]](#) This laboratory-based test can be completed in approximately 18 minutes.
- **GS HIV Combo Ag/Ab EIA:** This semi-automated laboratory assay can detect HIV-1 p24 antigen, antibodies to HIV-1 (groups O and M), and antibodies to HIV-2.[\[39\]](#) A reactive test does not distinguish p24 antigen from antibodies to HIV, and does not distinguish HIV-1 from HIV-2. In a performance evaluation, the GS HIV Combo Ag/Ab EIA was shown to be 100% sensitive in detecting established, previously confirmed HIV and more than 85% sensitive in detecting acute HIV infection, with specificity above 99% in both groups.[\[40\]](#) This test uses an EIA microwell format, and it takes at least 3 hours to perform. This is one of the only FDA-approved, laboratory-based, antigen-antibody assays considered high complexity by CLIA. It is semi-automated (whereas the others are fully automated); it can be performed manually but is more labor-intensive and has a longer turnaround time as compared to other antigen-antibody assays performed in the laboratory.
- **Liaison XL MUREX HIV Ag/Ab HT:** This chemiluminescent immunoassay, which is also described as a double-sandwich immunoassay, has high sensitivity and specificity for detecting p24 antigen, HIV-1 antibodies, and HIV-2 antibodies.[\[41\]](#) Testing can be performed with serum or plasma samples. This test, however, does not distinguish between the reactivity of p24 antigen and HIV-1 or HIV-2 antibodies.[\[41\]](#) The laboratory-based test requires approximately 32 minutes.
- **VITROS HIV Combo Test:** This laboratory assay can detect HIV-1 p24 antigen, antibodies to HIV-1 (groups O and M), and antibodies to HIV-2; this assay is part of the VITROS 3600 Immunodiagnostic System.[\[42,43\]](#) A reactive test does not distinguish p24 antigen from antibodies to HIV, and it does not distinguish HIV-1 from HIV-2. This assay is an immunometric 2-stage reaction, and it takes 48 minutes to perform.

A CDC study evaluated the reactivity of HIV tests in specimens from individuals with HIV-1 seroconversion and showed that an HIV-1/2 antigen-antibody immunoassay was positive in 50% of persons at 17.8 days and in 99% at 44.3 days ([Figure 8](#)).[\[28\]](#) Accordingly, the CDC considers laboratory-based HIV-1/2 antigen-antibody immunoassays to virtually exclude HIV infection if the test is negative 45 days after an exposure.[\[22\]](#)

HIV Antigen-Antibody Single-Use Point-of-Care Tests

In the United States, there is only one FDA-approved point-of-care HIV-1/2 antigen-antibody test for the diagnosis of HIV:

- **Abbott Determine HIV-1/2 Ag/Ab Combo:** This assay is a point-of-care, single-use, rapid test that

can detect HIV-1 p24 antigen, antibodies to HIV-1, and antibodies to HIV-2.[44,45] The Abbott Determine HIV-1/2 Ag/Ab Combo assay is the only FDA-approved point-of-care HIV-1/2 antigen-antibody test, and it is considered waived by CLIA when used for fingerstick whole blood. This assay can differentiate HIV-1 p24 antigen from HIV antibody, but it does not differentiate HIV-1 and HIV-2 antibodies.[46] The sensitivity of this assay for acute or very recent HIV infection is less than with laboratory-based HIV-1/2 antigen-antibody assays.[47,48,49] With this assay, the use of fingerstick whole blood specimens is not as sensitive as with plasma samples.[50] The Abbott Determine HIV-1/2 Ag/Ab Combo is a lateral flow immunochromatographic assay that takes 20 minutes to perform.

HIV Antibody Laboratory-Based Tests

The HIV enzyme immunoassay (EIA) antibody test for HIV diagnosis was first licensed in the United States in 1985.[20] For more than 20 years, HIV antibody tests were widely used as the initial laboratory diagnosis test in the HIV testing algorithm. Since 2014, however, the use of HIV antibody tests as an initial screening test has been replaced by HIV antigen-antibody assays.[14,15] A reactive HIV antibody test always requires further confirmatory testing with another HIV assay.[14] Most current laboratory-based HIV antibody tests are IgM/IgG-sensitive assays (as opposed to IgG-only assays) and can detect HIV IgM antibodies at approximately 23 to 25 days after HIV acquisition.[21] The CDC considers these HIV antibody tests to have a window period of 90 days (even the IgM/IgG-sensitive assays); in this context, a negative HIV EIA test 90 days after a possible HIV exposure is considered to effectively rule out HIV acquisition from that exposure.[22]

HIV Antibody Single-Use, Point-of-Care Tests

Single-use, point-of-care HIV test kits have self-contained testing reagents and materials and typically can yield a test result within 40 minutes.[20,51,52] There are 7 FDA-approved, rapid, point-of-care tests that the CDC identifies as suitable for use in clinical and nonclinical settings.[51] Six of these tests detect antibodies to HIV-1 and HIV-2, and one detects antibodies only to HIV-1. These antibody test results are either reactive or nonreactive. Hence, none of the currently used point-of-care antibody tests can differentiate HIV-1 from HIV-2. Multiple reports have shown problems with false-negative and false-positive test results with the oral fluid point-of-care test. Single-use, point-of-care rapid antibody tests are less sensitive than laboratory-based antigen-antibody tests for the detection of early HIV.[32,53,54,55] All positive point-of-care HIV test results are considered to be a presumptive positive test and require further supplemental testing for confirmation of HIV.[14] Single-use, point-of-care testing is primarily used for testing (1) in emergency room encounters where follow-up might be problematic, (2) persons in labor who had no HIV testing performed during their pregnancy, (3) in an occupational exposure to HIV when immediate results may be needed, and (4) in other clinical settings where a low likelihood of follow-up for HIV test results is anticipated.[20,56,57] The following list summarizes the current FDA-approved single-use, point-of-care, rapid HIV tests (in alphabetical order):[24]

- **Chembio DPP HIV 1/2 Assay:** This IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[58] This assay can be performed on samples of oral fluid, fingerstick whole blood, venous whole blood, serum, or plasma. This immunochromatographic test utilizes the Dual Path Platform (DPP) and requires 10-15 minutes to perform with blood samples and 25-40 minutes for oral fluid.
- **Chembio HIV 1/2 STAT-PAK Assay:** This IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[59] Tests can be performed using fingerstick whole blood, venous whole blood, serum, or plasma samples. This immunochromatographic lateral-flow test requires 15 minutes to perform.
- **Chembio SURE CHECK HIV 1/2 Assay:** This IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[60] This test is approved for use with fingerstick whole blood, venous whole blood, serum, or plasma samples. This immunochromatographic lateral-flow test requires 15 minutes to perform.
- **INSTI HIV-1/HIV-2 Antibody Test:** This IgM/IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[61] This test is approved for use with fingerstick whole blood, venipuncture whole blood, and plasma samples. This flow-through immunoassay requires only 2 minutes to perform.

- **OraQuick ADVANCE Rapid HIV-1/2 Antibody Test:** This IgM/IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[62] This assay can be performed on samples of oral fluid, fingerstick whole blood, venous whole blood, or plasma. This lateral flow immunoassay requires 20-40 minutes to perform.
- **Reveal G4 Rapid HIV-1 Antibody Test:** This IgG-sensitive assay can detect antibodies to HIV-1.[63] The following samples are FDA-approved for testing: fingerstick whole blood, venipuncture whole blood, serum, and plasma. This vertical flow immunoassay requires less than 3 minutes to perform.
- **Uni-Gold Recombigen HIV-1/2:** This IgM/IgG-sensitive assay can detect antibodies to HIV-1 and/or HIV-2.[64] This test is indicated for use with fingerstick whole blood, venous whole blood, serum, or plasma samples. This lateral-flow immunoassay requires 15-20 minutes to perform.

HIV-1/2 Differentiation Assays

Although several tests can distinguish HIV-1 from HIV-2, only one is currently in use (the Geenius HIV-1/2 Supplemental Assay), which is approved by the FDA as an HIV-1/HIV-2 differentiation assay.[65,66] Differentiating HIV-1 and HIV-2 is important to avoid misclassification of HIV infection; studies have shown that the HIV-1 Western blot was erroneously interpreted as positive for HIV-1 in 46% to 85% of specimens from persons with HIV-2.[14]

- **Geenius HIV 1/2 Supplemental Assay:** The Geenius HIV 1/2 Supplemental Assay is a single-use, immunochromatographic test that functions both as an HIV confirmatory test and an HIV-1 and HIV-2 differentiation assay (Figure 9).[65,67] This assay utilizes multiple recombinant or synthetic peptides to detect HIV-1 antibodies (p31, gp160, p24, and gp41) and HIV-2 antibodies (gp36 and gp140). The test cassette contains 7 test lines, including the 6 HIV peptides and one control. A positive HIV-1 result requires at least 2 envelope peptides (gp160 and gp41) or 1 envelope peptide plus either the p24 or the polymerase peptide p31; a positive HIV-2 diagnosis requires reactivity to both HIV-2 envelope peptides gp36 and gp140.[67] Investigators have shown the Geenius HIV 1/2 Supplemental Assay is a reliable HIV confirmatory assay, and this test is recommended by the CDC as the differentiation assay to use in the HIV testing algorithm.[68,69,70,71,72]. The result may indicate reactivity for HIV-1, reactivity for HIV-2, or reactivity for both in cases of HIV-1/HIV-2 coinfection. It may be run on serum, plasma, or whole blood.

HIV-1 Western Blot Laboratory Tests

The HIV-1 Western blot has been largely replaced by more sensitive and specific HIV diagnostic tests. When used, the HIV-1 Western blot can detect human antibodies that react to HIV-1 proteins that originate from three HIV-1 gene regions: *env* (gp41, gp120/160), *pol* (p31, p51, p66), and *gag* (p15, p17, p24, p55) (Figure 10).[73] The HIV Western blot typically becomes positive after about 5 to 6 weeks following HIV acquisition; as more protein bands become detectable, the Western blot typically evolves from a pattern of negative, then indeterminate, then positive.[73]

HIV SELF TESTING

When HIV testing is performed by the person undergoing testing, it is referred to as self testing or in-home testing. With the OraQuick In-Home HIV Test, the testing is done in its entirety at home (or in another private location). With the Home Access HIV-1 Test System, the specimen is collected at home (or in another private location) and mailed in for testing. Both of these commercially available tests provide individuals with an option for anonymous HIV testing. Self-testing may be preferable for some people who are reluctant to undergo HIV testing in medical settings.[74,75] Studies have shown that self-testing is feasible and acceptable for persons undergoing testing,[75,76,77,78,79] though several concerns persist, including (1) the cost of the test, (2) low sensitivity for detecting recent HIV acquisition, (3) lack of appropriate counseling and confirmatory testing for a positive test result, and (4) insufficient resources for linkage to care for persons with a positive test or linkage to HIV PrEP services for people with a negative test result.

- **OraQuick In-Home HIV Test:** The OraQuick In-Home HIV Test is the only FDA-approved HIV self test that includes specimen collection and interpretation of the HIV test results.[26] The test involves collecting an oral sample with a test device, placing the test device in a test kit vial that contains a developer solution, and then waiting 20 minutes to read the test result (the test must be read within 40 minutes). The client must read and interpret the test result. The test costs approximately \$40, and it includes a full set of easy-to-follow instructions. As with all other rapid tests, a positive in-home HIV test result is considered a preliminary positive HIV test result and confirmatory HIV testing is required. This test is offered by some public health programs for screening in other non-clinic settings for individuals who may not access healthcare and clinic-based testing options.

HIV Nucleic Acid Diagnostic Laboratory Tests

In the United States, HIV nucleic acid tests, including qualitative HIV RNA and quantitative HIV RNA assays, are used as part of the HIV diagnostic algorithm.[15] Given the very low limit of detection of most HIV quantitative HIV RNA assays, many clinicians now use quantitative HIV RNA tests (also known as viral load tests), rather than the FDA-approved qualitative HIV RNA assays, for diagnostic purposes. The quantitative tests are more widely available than the qualitative tests, as they are routinely used in the clinical management of persons with established HIV.[20,80] For routine HIV screening, the HIV NAT tests are not typically used due to high cost, technical complexity, and the failure to detect HIV elite controllers (the approximately 0.5% of individuals with HIV who maintain undetectable HIV RNA levels without antiretroviral therapy).[81] Instead, the HIV RNA tests are primarily used for confirmatory purposes. The HIV NAT is sometimes used for non-confirmatory purposes, including in persons being evaluated for acute HIV and persons receiving HIV PrEP (either oral or long-acting injectable HIV PrEP).[82] In this setting, the HIV-1 NAT is often used in combination with HIV-1/2 antigen-antibody testing to (1) exclude HIV prior to initiating HIV PrEP and (2) to assist in the diagnosis of HIV in persons receiving HIV PrEP, since persons who acquire HIV while taking HIV PrEP can have negative or ambiguous HIV screening test results if HIV-1/2 antigen-antibody testing alone.[82]

HIV nucleic acid testing may be used in four situations as an HIV diagnostic test:

- In the CDC/APHL HIV diagnostic algorithm to evaluate for possible acute infection when a specimen has a reactive HIV-1/2 antigen-antibody immunoassay, but a nonreactive or indeterminate HIV-1/HIV-2 differentiation assay.
- When a high suspicion of acute HIV exists and the initial HIV-1/2 antigen-antibody immunoassay result is negative. In this scenario, a negative HIV-1/2; antigen-antibody immunoassay and a positive HIV NAT confirms the diagnosis of acute (primary) HIV.
- To confirm chronic HIV-1 infection. Quantitative HIV RNA NAT is used routinely in clinical practice for monitoring the viral loads of patients who have already been diagnosed with HIV, and many commercially available tests are capable of detecting viremia as low as 20 copies/mL.
- As part of screening for HIV infection, in addition to an antigen-antibody assay, for persons planning to start or receiving HIV PrEP.

In the United States, there is currently only one HIV RNA NAT that is FDA-approved for HIV diagnostic purposes:

- **APTIMA HIV-1 RNA Qualitative Assay:** This laboratory-based, instrumented nucleic acid test is the only FDA-approved NAT for the diagnosis of HIV-1 infection, including acute HIV-1 infection.[83] This assay can detect all major HIV-1 groups and subtypes and has a manufacturer reported detection rate of 98.5% for 30 copies/mL, 82.6% for 10 copies/mL, and 42.5% for 3 copies/mL.[84,85] The APTIMA HIV-1 RNA Qualitative Assay requires at least 3 hours to perform.

Laboratory HIV Testing Algorithm as Recommended by CDC/APHL

The CDC and APHL HIV testing algorithm, which was initially published in 2014 and then updated in 2018, utilizes an HIV-1/2 antigen-antibody immunoassay as the initial test, with positive test results followed by an HIV-1/2 differentiation assay ([Figure 11](#)).[\[14,15,68\]](#) This HIV testing algorithm provides for a more accurate diagnosis of acute HIV-1, a more accurate diagnosis of HIV-2, fewer indeterminate results (due to a shorter window period), and a faster turnaround time than previous approaches.[\[14,15\]](#) Although the use of this algorithm will enhance earlier detection of acute HIV-1 infection, no single test is capable of detecting HIV immediately following HIV acquisition during the eclipse phase.[\[86\]](#) The rationale for using an algorithm that differentiates HIV-1 versus HIV-2 is that HIV-2 infection, though uncommon in the United States, significantly differs from HIV-1 with respect to natural history, the type of RNA assay (viral load) test needed for monitoring on antiretroviral treatment, and response to certain classes of antiretroviral medications, particularly non-nucleoside reverse transcriptase inhibitors (NNRTIs). From a practical standpoint, the same patient blood sample can be used for the initial screening test and the HIV differentiation assay. When using point-of-care sampling as the initial screening test, the confirmatory testing requires obtaining an additional sample.

Initial Testing

The recommended initial HIV test should be a laboratory-based HIV-1/2 antigen-antibody immunoassay; these tests can detect antibodies to HIV-1, antibodies to HIV-2, and HIV-1 p24 antigen.[\[14,15\]](#) A positive HIV-1/2 antigen-antibody immunoassay requires confirmation and differentiation of HIV-1 from HIV-2 infection. A person with a negative initial HIV-1/2 antigen-antibody immunoassay is considered to not have HIV infection, as long as a very recent (within approximately 4 weeks) exposure to HIV has not occurred. If no recent exposure to HIV has occurred, further HIV testing is not required for evaluation of current HIV status. In situations where it is not feasible to perform a laboratory-based initial HIV-1/2 antigen-antibody immunoassay, the rapid, point-of-care Determine HIV-1/2 Ag/Ab Combo test can be used with serum or plasma samples as the initial test in the HIV diagnostic laboratory algorithm. Note that the Determine HIV-1/2 Ag/Ab Combo is not as sensitive as the laboratory-based HIV-1/2 antigen-antibody immunoassays for detecting HIV during acute infection.[\[87\]](#)

Differentiation Assay

If the initial screening HIV-1/2 antigen-antibody immunoassay is reactive, a second HIV test is needed to confirm the initial test and to differentiate whether the infection is caused by HIV-1, HIV-2, or both. For this purpose, the CDC algorithm recommends using an HIV-1/HIV-2 antibody differentiation assay. The Geenius HIV 1/2 Supplemental Assay is the only FDA-approved assay currently in use for differentiating HIV-1 from HIV-2 infection.[\[68,69,88\]](#) Samples that are reactive with the HIV-1/2 antigen-antibody immunoassay and the HIV differentiation assay are considered positive and should be classified as HIV-1, HIV-2, or HIV-1 and HIV-2.

- Specimens that are reactive on the initial HIV-1/2 antigen-antibody immunoassay but either indeterminate or nonreactive on the differentiation assay require further testing with an HIV-1 NAT (qualitative or quantitative RNA assay) to evaluate the possibility of acute HIV (false-negative differentiation assay due to the window period) versus a false-positive HIV-1/2 antigen-antibody test. In this situation, if the NAT is positive for HIV-1, the person is likely to have acute HIV-1 infection.
- With the Geenius HIV-1/2 differentiation assay, the HIV-1 result can be positive, negative, or indeterminate, and the HIV-2 result can also be positive, negative, or indeterminate.[\[68\]](#) This gives a number of possible combinations of positive, negative, and indeterminate for the final result, all of which must be interpreted in the context of the pretest probability of HIV-1 or HIV-2 infection for a particular individual.[\[68\]](#)
- In the case of a reactive (“preliminary positive”) result from a rapid test, the specimen should be submitted for testing according to the full 2014 algorithm, beginning with the combination of HIV-1/2 antigen-antibody immunoassay.[\[14\]](#)

HIV Nucleic Acid Testing

If the initial HIV-1/2 antigen-antibody immunoassay is positive, but the HIV-1/HIV-2 differentiation assay is negative, further testing with an HIV-1 NAT should be performed. This is generally accomplished by drawing a sample for a qualitative or quantitative HIV-1 RNA assay, unless the individual has a substantial risk for HIV-2 infection or known exposure to HIV-2, in which case an HIV-2 RNA assay should be added. If both the HIV-1/2 antigen-antibody immunoassay and the HIV-1/HIV-2 differentiation assay are positive, then quantitative HIV RNA testing (viral load) is indicated—HIV-1 quantitative or HIV-2 quantitative, depending on whether HIV-1 or HIV-2 is identified on the differentiation assay.

Interpretation of Test Results

- If the HIV-1/2 antigen-antibody immunoassay is nonreactive, then the interpretation is no infection with HIV-1 or HIV-2, unless the individual undergoing testing has acquired HIV within the past 30 days. If acute HIV is suspected, then perform an HIV-1 RNA test.
- If the HIV-1/2 antigen-antibody immunoassay is reactive and the HIV-1/HIV-2 differentiation assay result is reactive for HIV-1 and nonreactive for HIV-2, then conclude the person has HIV-1 infection.
- If the HIV-1/2 antigen-antibody immunoassay is reactive and the HIV-1/HIV-2 differentiation assay result shows nonreactive HIV-1 and reactive HIV-2, then conclude the patient has HIV-2 infection.
- If the HIV-1/2 antigen-antibody immunoassay is reactive and the HIV-1/HIV-2 differentiation assay result shows reactive HIV-1 and reactive HIV-2, then conclude the patient has HIV-1 and HIV-2 coinfection.
- If the HIV-1/2 antigen-antibody immunoassay is reactive and the HIV-1/HIV-2 differentiation assay shows HIV-1 indeterminate (or negative) in conjunction with a nonreactive HIV-2, then several possibilities exist. In this scenario, follow-up testing with HIV-1 RNA is indicated. If the HIV-1 RNA is positive, the patient has acute HIV-1. If the HIV-1 RNA is negative, the most probable scenario is that the initial reactive immunoassay result was a false-positive result, and the individual undergoing testing does not likely have HIV-1 or HIV-2. Alternatively, in a person with risk factors for acquiring HIV-2, these test results could theoretically indicate acute HIV-2. Follow-up testing with HIV-2 NAT should be considered if an individual has epidemiologic risk factors for exposure to HIV-2.

Performance of Diagnostic Tests

Characteristics of an Ideal Screening Test

The principles that define a good screening test are not unique to HIV infection and apply to medical screening in general. An ideal screening test will accurately identify individuals with the clinical condition of interest, but without mistakenly diagnosing individuals who do not have the condition. In addition, the use of screening tests is most effective when limited to conditions for which there exists available, effective treatment that can directly target the disease and improve prognosis and outcomes.[89]

Sensitivity

In relation to HIV testing, sensitivity refers to the proportion of true positives (persons who have HIV infection) that are correctly identified by a screening test (Figure 12).[90] In general, very high sensitivity is desired for initial HIV screening tests since the goal of the screening test is to not miss detecting anyone who has HIV infection. Thus, if the test is 100% sensitive and the person tests negative, you can be confident the individual tested does not have the infection. For example, all HIV antibody tests approved for use in the United States have a sensitivity greater than 98% for diagnosing persons with chronic HIV.[18]

Specificity

Specificity is the proportion of true negative persons who do not have HIV and are correctly identified as HIV-negative by a screening test (Figure 13).[91] If a test is 100% specific and the person tests positive, you can be confident they have the infection and the test is not a false-positive result. In the United States, initial HIV antigen-antibody tests have greater than 99% specificity for chronic HIV infection, and the specificity increases to nearly 100% when the initial test is combined with a supplemental HIV test, such as an HIV differentiation assay.[70]

Positive Predictive Value and Negative Predictive Value

In contrast to sensitivity and specificity, which refer to the diagnostic ability of a screening test, the predictive value of a test refers to the likelihood that the test will give the correct diagnosis.[90] Positive predictive value is the proportion of patients with a positive HIV result who are correctly diagnosed (i.e., who actually have HIV). Negative predictive value is the proportion of patients with negative HIV results who are correctly diagnosed (i.e., who truly do not have HIV).[90] Because screening tests are neither 100% sensitive nor 100% specific, the predictive value of tests is also imperfect. It is possible for a person to receive an incorrect result from a diagnostic test (these results are termed false-negative and false-positive test results). It is important to understand that the prevalence of a disease in a community impacts the predictive value of a given test, and predictive values in one study or in one community do not apply to all other settings.[90]

False-Negative HIV Tests

A false-negative HIV test result refers to a negative HIV test result in a person who actually has HIV (Figure 14). A false-negative HIV antibody (or antigen-antibody) test result most often occurs when performing testing in a person with acute HIV, from laboratory error, or following receipt of potent antiretroviral therapy very early after HIV acquisition.[92,93,94,95] In addition, rare causes of false-negative results include (1) persons who have defects in HIV-specific immunity and thus fail to generate certain antibodies [96,97,98,99], (2) persons who have acquired HIV while receiving HIV PrEP,[100,101] (3) persons with hypogammaglobulinemia,[102] and (4) persons who recently received potent immunosuppressant medications.[103] In adults with chronic HIV, the loss of HIV antibodies (seroreversion) is exceedingly rare.[104] A false-negative HIV p24 antigen test can occur in the first several weeks after HIV acquisition (usually this test is positive by day 17); in addition, many persons with untreated chronic HIV do not have persistently detectable p24 antigen levels, often due to p24 antigen complexing with p24 antibody. A false-

negative HIV NAT can occur in the first week or two after HIV acquisition during the eclipse phase (this test is typically positive on about day 10) and in the rare persons with chronic HIV who inherently have very strong immunologic control of HIV (elite controllers) and thus may have undetectable HIV RNA levels in the absence of antiretroviral therapy.

False-Positive HIV Tests

A false-positive HIV test result is defined as a positive HIV test result in a person who does not have HIV ([Figure 15](#)). A false-positive HIV test may occur due to polyclonal cross-reactivity, which is more common in the setting of pregnancy, recent inoculation with influenza vaccine (or other vaccines), autoimmune disorders, receipt of an investigational HIV-1 vaccine, receipt of gamma globulin, prior blood transfusions, human T-lymphotropic virus (HTLV) infection, recent viral infection (including COVID-19), collagen vascular diseases, and laboratory errors.[\[105,106\]](#) Recently, several reports have described false-positive HIV NATs in persons who received chimeric antigen receptor (CAR) T-cell therapy, due to the lentivirus used as the vector in manufacturing these individualized therapies; in these cases, the lentivirus vector used had incorporated a transgene plasmid that contained part or all of the HIV *gag* sequence.[\[107\]](#) When trying to determine whether a person's HIV screening test result is accurate, the pretest probability—the likelihood before the test was performed that the patient has HIV—can help with interpretation. Further, the likelihood of an accurate HIV test result correlates directly with the prevalence of HIV in the testing community: the proportion of false-positive tests is higher in populations with low HIV prevalence (even if the screening test is highly sensitive and specific), whereas the proportion of false-negative tests is lower.[\[90\]](#)

Special Diagnostic Situations

Diagnosis of Acute HIV-1

The laboratory diagnosis of acute HIV-1 infection is most reliably made with a positive HIV RNA (or HIV-1 p24 antigen) with a concomitant negative HIV antibody assay; note that with very early acute HIV infection, the p24 antigen assay may be negative ([Figure 16](#)).[\[17,108,109\]](#) Use of HIV-1/2 antigen-antibody immunoassays will detect HIV about 17 days after HIV acquisition, which is significantly sooner than with HIV laboratory-based HIV antibody tests, all point-of-care HIV tests, and in-home HIV tests.[\[19,86,110,111\]](#) Even when using HIV-1/2 antigen-antibody immunoassays, the initial laboratory testing will fail to detect some individuals who have very early acute HIV infection. Thus, for individuals in whom initial HIV-1/2 antigen-antibody testing is nonreactive, but acute HIV is strongly suspected, HIV NAT (i.e., HIV RNA testing) should be performed. Increased awareness of acute retroviral syndrome by medical providers can help facilitate diagnosis in the early stages of infection. Among persons recently infected with HIV, it is estimated that at least half develop a nonspecific syndrome characterized by fever, myalgia, lymphadenopathy, pharyngitis, fatigue, headache, and rash (a mononucleosis-like syndrome).[\[112,113,114\]](#) Because HIV RNA levels are typically very high in persons with acute retroviral syndrome, an HIV NAT is uniformly positive at this stage of infection. Ideally, if acute HIV is suspected, a quantitative RNA assay (as opposed to a qualitative RNA assay) should be ordered since HIV RNA levels are typically very high at this stage.

Diagnosing HIV In Persons Receiving HIV PrEP

The diagnostic accuracy and timing of early HIV infection in persons who acquire HIV while taking HIV PrEP with either tenofovir DF-emtricitabine, tenofovir alafenamide-emtricitabine, or long-acting injectable cabotegravir, may result in atypical laboratory patterns, such as delayed seroconversion, indeterminate results on HIV differentiation assays, low-level HIV RNA levels, or HIV RNA levels below the limit of the assay, even in persons with acute or early HIV.[\[115\]](#) Data from the Partners PrEP Study and the Bangkok Tenofovir Study showed persons receiving HIV PrEP who acquired HIV had marked delays in HIV seroconversion with point-of-care tests, especially when using oral fluid samples.[\[100,116\]](#) Other groups have reported false-negative or ambiguous HIV test results in persons taking HIV PrEP.[\[101,115\]](#) Problems with false-negative testing are greater in this setting when using point-of-care tests; thus, laboratory-based HIV testing is recommended when monitoring persons receiving HIV PrEP.[\[21\]](#) Of note, even laboratory-based, antigen-antibody tests may be affected by HIV PrEP exposure. In the HPTN 083 trial, which compared injectable cabotegravir to daily, oral tenofovir DF-emtricitabine, delays in diagnosis were documented in individuals with incident HIV acquisition in both arms, though delays were generally longer for individuals in the cabotegravir arm.[\[117\]](#) For this reason, the CDC HIV PrEP Guidelines, which were updated in 2021, now recommend routine HIV RNA monitoring for individuals receiving HIV PrEP, and an HIV RNA test prior to initiating HIV PrEP is recommended in several circumstances.[\[82\]](#) In situations where results are ambiguous or confusing, clinical consultation is recommended.

Diagnosis of HIV in Infants and Children Exposed to HIV

The 2014 and 2018 CDC HIV diagnostic algorithm does not address HIV diagnostic testing of infants and children exposed to HIV.[\[14,15\]](#) To diagnose HIV among infants younger than 18 months of age, the Pediatric ART Guidelines recommend using a virologic assay (HIV NAT) that directly detects HIV RNA or HIV DNA (either quantitative or qualitative tests can be used). This issue is addressed in detail in the section Diagnosis of HIV in Infants and Children in the lesson [HIV in Infants and Children](#).

Diagnosis of HIV-2

The 2014 and 2018 HIV diagnostic algorithm improves the detection of HIV-2 by using an HIV-1/HIV-2 differentiation assay as the second step of the algorithm (following the initial HIV-1/2 antigen-antibody immunoassay).[\[14,15,118\]](#) Confirmation of HIV-2 infection can be challenging since HIV-1 RNA assays do not

reliably detect or quantitate HIV-2. More recently, quantitative HIV-2 RNA assays have become available through the University of Washington Department of Laboratory Medicine ([HIV-2 RNA Quantitation](#)) and the New York State Department of Health ([HIV-2 Nucleic Acid Testing](#), which includes HIV-2 qualitative or quantitative options).^[119] It is important to note, however, that a significant percentage of individuals with HIV-2 have undetectable HIV-2 RNA levels without antiretroviral treatment. Thus, in certain epidemiological settings (e.g., a person with risk factors for acquiring HIV-2), a positive screening HIV-1/2 antigen-antibody test followed by a positive HIV-2 antibody on the differentiation assay should be considered HIV-2 positive even if plasma HIV-2 RNA is undetectable with an HIV-2 RNA assay. Prior to 2014, the diagnosis of HIV-2 was often missed or delayed due to improper classification as HIV-1.^[120] This occurred because an HIV Western blot was used as the confirmatory test (instead of the currently used HIV-1/2 differentiation assay), and HIV-2 infection may cause a negative, indeterminate, or positive HIV-1 Western blot due to cross-reacting antibodies.^[14,120,121]

Delivering Test Results

Follow-Up for Test Results

The use of multiple modalities for HIV testing and delivery of HIV test results has helped to optimize this process. In particular, delivery of HIV test results by telephone has been found to be both effective and acceptable, and increases the number of people who receive their test results.[122,123] In addition, the use of point-of-care, rapid tests, which provide a result at the same visit and augment the capacity to run tests quickly at mobile and community sites, provides an opportunity for real-time delivery of test results.[124] Implementation of rapid tests in outreach and community settings was found in this report to enhance testing of people at risk of acquiring HIV but who may not regularly have access to medical care. The availability of self-testing also increases the proportion of persons who undergo HIV testing and can immediately see their test result. That said, a positive self-test result always requires confirmatory testing and recent large studies found mixed results when assessing whether self-testing significantly reduces the number of individuals with undiagnosed HIV.[77,125,126]

Communicating Test Results

The CDC offers practical advice for medical providers who offer HIV testing in their clinical settings.[127] Medical providers should be prepared to deliver results to individuals undergoing HIV testing in a private area and in a direct, neutral tone. The health care professional delivering the test results should be knowledgeable about HIV, since persons undergoing testing may have questions about HIV, risk of transmission to partners, and disclosure of HIV status to partners. Any individual who receives a positive HIV test result should be linked to HIV care prior to leaving the testing setting and should have a scheduled appointment with an HIV medical provider as soon as possible. Availability of a case manager or social worker familiar with HIV and HIV-related resources can aid in the initial discussion with a person newly diagnosed with HIV. Being able to provide emotional support, medical information, and timely linkage to care is critical when delivering positive HIV results.[128] For persons who test negative for HIV, the medical provider should be prepared to provide HIV prevention counseling to help the individual remain HIV negative, including discussion of and referral for HIV PrEP, if indicated.

Status Neutral Approach for HIV Prevention and Care

The CDC recommends a status neutral approach to HIV care, which means that clinics and other healthcare sites offer culturally-sensitive, stigma-free care to all individuals, encompassing HIV testing, prevention, and treatment.[13] This approach begins with an HIV test, as recommended for all adults and adolescents in the United States, and involves offering HIV preventative resources and care for those who test negative and have a risk of acquiring HIV, as well as offering HIV treatment for individuals who have a confirmed HIV positive test.

Summary Points

- Laboratory markers of HIV infection (HIV RNA, p24 antigen, anti-HIV IgM antibody, anti-HIV IgG antibody) appear in a consistent sequence and are the basis for all of the HIV diagnostic tests.
 - In 2014, the Centers for Disease Control and Prevention (CDC) and the American Public Health Laboratories (APHL) jointly published new HIV diagnostic testing guidelines.
 - The CDC HIV testing algorithm recommends initial testing with an HIV-1/2 antigen-antibody immunoassay, followed (for samples that are reactive on the antigen-antibody test) by an HIV-1/2 differentiation assay. Testing for HIV RNA should be done in cases where the initial test is reactive, but the differentiation assay is either nonreactive or indeterminate.
 - Compared to previous screening algorithms, the current algorithm is (1) more likely to detect acute HIV-1, (2) more accurately diagnoses HIV-2, (3) allows for faster turnaround time, and (4) leads to fewer indeterminate results.
 - An ideal screening test is sensitive, specific, and limited to conditions for which there is effective treatment available that can directly target the disease and improve prognosis and outcomes.
 - False-negative HIV screening test results can occur during acute HIV; false-positive HIV screening test results may occur due to laboratory errors, and rarely from cross-reactivity with other antibodies, such as during pregnancy, in persons who have an autoimmune condition, or following recent vaccine administration.
 - Testing for HIV RNA may identify very early HIV infection (HIV RNA tests may be positive up to a week sooner than the antigen-antibody tests), but HIV RNA is typically not detected in the first 10 days after induction during the eclipse phase.
 - Single-use, point-of-care HIV tests and self-testing are additional options to help facilitate HIV screening and detection. A reactive result on a point-of-care or self-test should be considered a presumptive positive result and requires further testing.
 - Challenges may occur with the diagnostic evaluation of acute HIV, when evaluating infants and children exposed to HIV, and in persons receiving HIV PrEP. Clinical consultation is recommended if a person has been exposed to HIV or there is suspicion or risk for HIV, yet HIV test results are ambiguous or indeterminate.
-
- Diagnostic testing for HIV should ideally be performed as part of status neutral care, which involves delivering culturally-sensitive, stigma-free care to all individuals and linking persons who test negative to appropriate preventive care and those who test positive to HIV treatment services.

Citations

1. Cheever LW. Engaging HIV-infected patients in care: their lives depend on it. *Clin Infect Dis*. 2007;44:1500-2.
[PubMed Abstract] -
2. El-Sadr WM, Lundgren J, Neaton JD, et al. CD4+ count-guided interruption of antiretroviral treatment. *N Engl J Med*. 2006;355:2283-96.
[PubMed Abstract] -
3. Lundgren JD, Babiker AG, Gordin F, et al. Initiation of Antiretroviral Therapy in Early Asymptomatic HIV Infection. *N Engl J Med*. 2015;373:795-807.
[PubMed Abstract] -
4. TEMPRANO ANRS 12136 Study Group, Danel C, Moh R, et al. A Trial of Early Antiretrovirals and Isoniazid Preventive Therapy in Africa. *N Engl J Med*. 2015;373:808-22.
[PubMed Abstract] -
5. INSIGHT START Study Group, Lundgren JD, Babiker AG, Gordin F, et al. Initiation of Antiretroviral Therapy in Early Asymptomatic HIV Infection. *N Engl J Med*. 2015;373:795-807.
[PubMed Abstract] -
6. Cohen MS, Chen YQ, McCauley M, et al. Antiretroviral Therapy for the Prevention of HIV-1 Transmission. *N Engl J Med*. 2016;375:830-9.
[PubMed Abstract] -
7. Li Z, Purcell DW, Sansom SL, Hayes D, Hall HI. Vital Signs: HIV transmission along the continuum of care - United States, 2016. *MMWR Morb Mortal Wkly Rep*. 2019;68:267-72.
[PubMed Abstract] -
8. Prabhu VS, Hutchinson AB, Farnham PG, Sansom SL. Sexually acquired HIV infections in the United States due to acute-phase HIV transmission: an update. *AIDS*. 2009;23:1792-4.
[PubMed Abstract] -
9. Pinkerton SD. How many sexually-acquired HIV infections in the USA are due to acute-phase HIV transmission? *AIDS*. 2007;21:1625-9.
[PubMed Abstract] -
10. Pilcher CD, Tien HC, Eron JJ Jr, et al. Brief but efficient: acute HIV infection and the sexual transmission of HIV. *J Infect Dis*. 2004;189:1785-92.
[PubMed Abstract] -
11. Hollingsworth TD, Anderson RM, Fraser C. HIV-1 transmission, by stage of infection. *J Infect Dis*. 2008;198:687-93.
[PubMed Abstract] -
12. Centers for Disease Control and Prevention. Estimated HIV Incidence and Prevalence in the United States, 2018–2022. HIV Surveillance Supplemental Report. 2024;29(No. 1):1-131. Published May 2024.
[CDC] -
13. Centers for Disease Control and Prevention. Issue Brief: Status Neutral HIV Care and Service Delivery Eliminating Stigma and Reducing Health Disparities.
[CDC] -

14. Centers for Disease Control and Prevention and Association of Public Health Laboratories. Laboratory Testing for the Diagnosis of HIV Infection: Updated Recommendations. Published June 27, 2014. [\[CDC\]](#) -
15. Centers for Disease Control and Prevention and Association of Public Health Laboratories. 2018 Quick reference guide: Recommended laboratory HIV testing algorithm for serum or plasma specimens. Published January 27, 2018. [\[CDC\]](#) -
16. Fiebig EW, Wright DJ, Rawal BD, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. *AIDS*. 2003;17:1871-9. [\[PubMed Abstract\]](#) -
17. Panel on Antiretroviral Guidelines for Adults and Adolescents. Guidelines for the use of antiretroviral agents in adults and adolescents with HIV. Department of Health and Human Services. Considerations for antiretroviral use in special patient populations: early (acute and recent) HIV infection. December 6, 2023. [\[HIV.gov\]](#) -
18. Masciotra S, McDougal JS, Feldman J, Sprinkle P, Wesolowski L, Owen SM. Evaluation of an alternative HIV diagnostic algorithm using specimens from seroconversion panels and persons with established HIV infections. *J Clin Virol*. 2011;52 Suppl 1:S17-22. [\[PubMed Abstract\]](#) -
19. Owen SM, Yang C, Spira T, et al. Alternative algorithms for human immunodeficiency virus infection diagnosis using tests that are licensed in the United States. *J Clin Microbiol*. 2008;46:1588-95. [\[PubMed Abstract\]](#) -
20. Branson BM. State of the art for diagnosis of HIV infection. *Clin Infect Dis*. 2007;45 Suppl 4:S221-5. [\[PubMed Abstract\]](#) -
21. Hurt CB, Nelson JAE, Hightow-Weidman LB, Miller WC. Selecting an HIV Test: A Narrative Review for Clinicians and Researchers. *Sex Transm Dis*. 2017;44:739-46. [\[PubMed Abstract\]](#) -
22. Delaney KP, Wesolowski LG, Owen SM. The Evolution of HIV Testing Continues. *Sex Transm Dis*. 2017;44:747-9. [\[PubMed Abstract\]](#) -
23. Branson BM, Mermin J. Establishing the diagnosis of HIV infection: new tests and a new algorithm for the United States. *J Clin Virol*. 2011;52 Suppl 1:S3-4. [\[PubMed Abstract\]](#) -
24. U.S. Food and Drug Administration. Complete List of Donor Screening Assays for Infectious Agents and HIV Diagnostic Assays. [\[FDA\]](#) -
25. Centers for Disease Control and Prevention. Advantages and disadvantages of FDA-approved HIV immunoassays used for screening by generation and platform. [\[CDC\]](#) -
26. U.S. Food and Drug Administration. OraQuick In-Home HIV Test. [\[U.S. FDA\]](#) -

27. Daskalakis D. HIV diagnostic testing: evolving technology and testing strategies. *Top Antivir Med.* 2011;19:18-22.
[\[PubMed Abstract\]](#) -
28. Delaney KP, Hanson DL, Masciotra S, Ethridge SF, Wesolowski L, Owen SM. Time Until Emergence of HIV Test Reactivity Following Infection With HIV-1: Implications for Interpreting Test Results and Retesting After Exposure. *Clin Infect Dis.* 2017;64:53-9.
[\[PubMed Abstract\]](#) -
29. U.S. Food and Drug Administration. ADVIA Centaur HIV Ag/Ab Combo (CHIV) Assay.
[\[U.S. FDA\]](#) -
30. Pumarola T, Freeman J, Saxton E, Dillon P, Bal T, van Helden J. Performance evaluation of the ADVIA Centaur(®) HIV Ag/Ab Combo assay. *J Virol Methods.* 2010;170:16-20.
[\[PubMed Abstract\]](#) -
31. U.S. Food and Drug Administration. Abbott ARCHITECT HIV Ag/Ab Combo.
[\[U.S. FDA\]](#) -
32. Stekler JD, Swenson PD, Coombs RW, et al. HIV testing in a high-incidence population: is antibody testing alone good enough? *Clin Infect Dis.* 2009;49:444-53.
[\[PubMed Abstract\]](#) -
33. Chavez P, Wesolowski L, Patel P, Delaney K, Owen SM. Evaluation of the performance of the Abbott ARCHITECT HIV Ag/Ab Combo Assay. *J Clin Virol.* 2011;52 Suppl 1:S51-5.
[\[PubMed Abstract\]](#) -
34. Pandori MW, Hackett J Jr, Louie B, Vallari A, Dowling T, Liska S, Klausner JD. Assessment of the ability of a fourth-generation immunoassay for human immunodeficiency virus (HIV) antibody and p24 antigen to detect both acute and recent HIV infections in a high-risk setting. *J Clin Microbiol.* 2009;47:2639-42.
[\[PubMed Abstract\]](#) -
35. U.S. Food and Drug Administration. BioPlex 2200 HIV Ag-Ab Assay.
[\[U.S. FDA\]](#) -
36. Salmona M, Delarue S, Delaugerre C, Simon F, Maylin S. Clinical evaluation of BioPlex 2200 HIV Ag-Ab, an automated screening method providing discrete detection of HIV-1 p24 antigen, HIV-1 antibody, and HIV-2 antibody. *J Clin Microbiol.* 2014;52:103-7.
[\[PubMed Abstract\]](#) -
37. U.S. Food and Drug Administration. Elecsys HIV combi PT.
[\[U.S. FDA\]](#) -
38. U.S. Food and Drug Administration. Elecsys HIV Duo.
[\[U.S. FDA\]](#) -
39. U.S. Food and Drug Administration. GS HIV Combo Ag/Ab EIA.
[\[U.S. FDA\]](#) -
40. Bentsen C, McLaughlin L, Mitchell E, et al. Performance evaluation of the Bio-Rad Laboratories GS HIV Combo Ag/Ab EIA, a 4th generation HIV assay for the simultaneous detection of HIV p24 antigen and antibodies to HIV-1 (groups M and O) and HIV-2 in human serum or plasma. *J Clin Virol.* 2011;52 Suppl

- 1:S57-61.
[\[PubMed Abstract\]](#) -
41. U.S. Food and Drug Administration. LIAISON XL MUREX HIV Ab/Ag HT and Control HIV Ab/Ag HT
[\[U.S. FDA\]](#) -
42. De Paschale M, Manco MT, Cianflone A, et al. Evaluation of the vitros hiv combo 4th generation test for the identification of HIV infections. J Clin Virol. 2018;108:77-82.
[\[PubMed Abstract\]](#) -
43. U.S. Food and Drug Administration. VITROS Immunodiagnostic Products HIV Combo Reagent Pack & VITROS Immunodiagnostic Products HIV Combo Calibrator.
[\[U.S. FDA\]](#) -
44. U.S. Food and Drug Administration. Alere Determine HIV-1/2 Ag/Ab Combo.
[\[U.S. FDA\]](#) -
45. Abbott Diagnostics. Determine HIV-1/2 Ag/Ab Combo.
[\[Abbott Diagnostics\]](#) -
46. Masciotra S, Luo W, Youngpairoj AS, et al. Performance of the Alere Determine™ HIV-1/2 Ag/Ab Combo Rapid Test with specimens from HIV-1 seroconverters from the US and HIV-2 infected individuals from Ivory Coast. J Clin Virol. 2013;58 Suppl 1:e54-8.
[\[PubMed Abstract\]](#) -
47. Rosenberg NE, Kamanga G, Phiri S, et al. Detection of acute HIV infection: a field evaluation of the determine® HIV-1/2 Ag/Ab combo test. J Infect Dis. 2012;205:528-34.
[\[PubMed Abstract\]](#) -
48. Duong YT, Mavengere Y, Patel H, et al. Poor performance of the determine HIV-1/2 Ag/Ab combo fourth-generation rapid test for detection of acute infections in a National Household Survey in Swaziland. J Clin Microbiol. 2014;52:3743-8.
[\[PubMed Abstract\]](#) -
49. Parker MM, Bennett SB, Sullivan TJ, et al. Performance of the Alere Determine™ HIV-1/2 Ag/Ab Combo Rapid Test with algorithm-defined acute HIV-1 infection specimens. J Clin Virol. 2018;104:89-91.
[\[PubMed Abstract\]](#) -
50. Masciotra S, Luo W, Westheimer E, et al. Performance evaluation of the FDA-approved Determine™ HIV-1/2 Ag/Ab Combo assay using plasma and whole blood specimens. J Clin Virol. 2017;91:95-100.
[\[PubMed Abstract\]](#) -
51. Centers for Disease Control and Prevention (CDC). HIV Testing Technologies: CLIA-Waived Rapid HIV Tests.
[\[CDC\]](#) -
52. Delaney KP, Branson BM, Uniyal A, et al. Performance of an oral fluid rapid HIV-1/2 test: experience from four CDC studies. AIDS. 2006;20:1655-60.
[\[PubMed Abstract\]](#) -
53. Pilcher CD, Louie B, Facente S, et al. Performance of rapid point-of-care and laboratory tests for acute and established HIV infection in San Francisco. PLoS One. 2013;8:e80629.
[\[PubMed Abstract\]](#) -

54. Stekler JD, O'Neal JD, Lane A, et al. Relative accuracy of serum, whole blood, and oral fluid HIV tests among Seattle men who have sex with men. *J Clin Virol.* 2013;58 Suppl 1:e119-22.
[\[PubMed Abstract\]](#) -
55. Stekler JD, Ure G, O'Neal JD, et al. Performance of Determine Combo and other point-of-care HIV tests among Seattle MSM. *J Clin Virol.* 2016;76:8-13.
[\[PubMed Abstract\]](#) -
56. Kuhar DT, Henderson DK, Struble KA, et al. Updated US Public Health Service Guidelines for the Management of Occupational Exposures to Human Immunodeficiency Virus and Recommendations for Postexposure Prophylaxis. *Infect Control Hosp Epidemiol.* 2013;34:875-92.
[\[PubMed Abstract\]](#) -
57. Centers for Disease Control and Prevention. Advancing HIV prevention: new strategies for a changing epidemic--United States, 2003. *MMWR Morb Mortal Wkly Rep.* 2003;52:329-32.
[\[PubMed Abstract\]](#) -
58. U.S. Food and Drug Administration. Chembio DPP HIV 1/2 Assay.
[\[U.S. FDA\]](#) -
59. U.S. Food and Drug Administration. Chembio HIV 1/2 STAT-PAK Assay.
[\[U.S. FDA\]](#) -
60. U.S. Food and Drug Administration. Chembio SURE CHECK HIV 1/2 Assay.
[\[U.S. FDA\]](#) -
61. U.S. Food and Drug Administration. INSTI HIV-1/HIV-2 Antibody Test.
[\[U.S. FDA\]](#) -
62. U.S. Food and Drug Administration. OraQuick ADVANCE Rapid HIV-1/2 Antibody Test.
[\[U.S. FDA\]](#) -
63. U.S. Food and Drug Administration. Reveal G4 Rapid HIV-1 Antibody Test (Reveal G4).
[\[U.S. FDA\]](#) -
64. U.S. Food and Drug Administration. Uni-Gold Recombigen HIV-1/2.
[\[U.S. FDA\]](#) -
65. U.S. Food and Drug Administration. Geenius HIV 1/2 Supplemental Assay.
[\[U.S. FDA\]](#) -
66. U.S. Food and Drug Administration. Multispot HIV-1/HIV-2 Rapid Test.
[\[U.S. FDA\]](#) -
67. Fernández McPhee C, Álvarez P, Prieto L, et al. HIV-1 infection using dried blood spots can be confirmed by Bio-Rad Geenius™ HIV 1/2 confirmatory assay. *J Clin Virol.* 2015;63:66-9.
[\[PubMed Abstract\]](#) -
68. Centers for Disease Control and Prevention (CDC). Technical Update on HIV-1/2 Differentiation Assays. August 12, 2016.
[\[CDC\]](#) -
69. Herssens N, Beelaert G, Franssen K. Discriminatory capacity between HIV-1 and HIV-2 of the new rapid confirmation assay Geenius. *J Virol Methods.* 2014;208:11-5.

[\[PubMed Abstract\]](#) -

70. Malloch L, Kadivar K, Putz J, et al. Comparative evaluation of the Bio-Rad Geenius HIV-1/2 Confirmatory Assay and the Bio-Rad Multispot HIV-1/2 Rapid Test as an alternative differentiation assay for CLSI M53 algorithm-I. J Clin Virol. 2013;58 Suppl 1:e85-91.
[\[PubMed Abstract\]](#) -
71. Montesinos I, Eykmans J, Delforge ML. Evaluation of the Bio-Rad Geenius HIV-1/2 test as a confirmatory assay. J Clin Virol. 2014;60:399-401.
[\[PubMed Abstract\]](#) -
72. Keating SM, Kassanje R, Lebedeva M, et al. Performance of the Bio-Rad Geenius HIV1/2 Supplemental Assay in Detecting "Recent" HIV Infection and Calculating Population Incidence. J Acquir Immune Defic Syndr. 2016;73:581-8.
[\[PubMed Abstract\]](#) -
73. Centers for Disease Control (CDC). Interpretation and use of the Western blot assay for serodiagnosis of human immunodeficiency virus type 1 infections. MMWR Morb Mortal Wkly Rep. 1989;38(Suppl 7):1-7.
[\[MMWR\]](#) -
74. Carballo-Diéguez A, Frasca T, Dolezal C, Balan I. Will gay and bisexually active men at high risk of infection use over-the-counter rapid HIV tests to screen sexual partners? J Sex Res. 2012;49:379-87.
[\[PubMed Abstract\]](#) -
75. Myers JE, El-Sadr WM, Zerbe A, Branson BM. Rapid HIV self-testing: long in coming but opportunities beckon. AIDS. 2013;27:1687-95.
[\[PubMed Abstract\]](#) -
76. Ricca AV, Hall EW, Khosropour CM, Sullivan PS. Factors Associated with Returning At-Home Specimen Collection Kits for HIV Testing among Internet-Using Men Who Have Sex with Men. J Int Assoc Provid AIDS Care. 2016;15:463-9.
[\[PubMed Abstract\]](#) -
77. Myers JE, Bodach S, Cutler BH, Shepard CW, Philippou C, Branson BM. Acceptability of home self-tests for HIV in New York City, 2006. Am J Public Health. 2014;104:e46-8.
[\[PubMed Abstract\]](#) -
78. Frank AP, Wandell MG, Headings MD, Conant MA, Woody GE, Michel C. Anonymous HIV testing using home collection and telemedicine counseling. A multicenter evaluation. Arch Intern Med. 1997;157:309-14.
[\[PubMed Abstract\]](#) -
79. Branson BM. Home sample collection tests for HIV infection. JAMA. 1998;280:1699-701.
[\[PubMed Abstract\]](#) -
80. Karris MY, Anderson CM, Morris SR, Smith DM, Little SJ. Cost savings associated with testing of antibodies, antigens, and nucleic acids for diagnosis of acute HIV infection. J Clin Microbiol. 2012;50:1874-8.
[\[PubMed Abstract\]](#) -
81. Madec Y, Boufassa F, Porter K, et al. Natural history of HIV-control since seroconversion. AIDS. 2013;27:2451-60.
[\[PubMed Abstract\]](#) -

82. Centers for Disease Control and Prevention: US Public Health Service: Preexposure prophylaxis for the prevention of HIV infection in the United States—2021 Update: a clinical practice guideline. December 2021;1-108.
[\[CDC\]](#) -
83. U.S. Food and Drug Administration. APTIMA HIV-1 RNA Qualitative Assay
[\[U.S. FDA\]](#) -
84. Giachetti C, Linnen JM, Kolk DP, et al. Highly sensitive multiplex assay for detection of human immunodeficiency virus type 1 and hepatitis C virus RNA. J Clin Microbiol. 2002;40:2408-19.
[\[PubMed Abstract\]](#) -
85. Pierce VM, Neide B, Hodinka RL. Evaluation of the Gen-Probe Aptima HIV-1 RNA qualitative assay as an alternative to Western blot analysis for confirmation of HIV infection. J Clin Microbiol. 2011;49:1642-5.
[\[PubMed Abstract\]](#) -
86. Branson BM, Stekler JD. Detection of acute HIV infection: we can't close the window. J Infect Dis. 2011;205:521-4.
[\[PubMed Abstract\]](#) -
87. Centers for Disease Control and Prevention (CDC). Technical Update: Use of the Determine HIV 1/2 Ag/Ab Combo Test with Serum or Plasma in the Laboratory Algorithm for HIV Diagnosis. October 4, 2017.
[\[CDC\]](#) -
88. Ramos EM, Harb S, Dragavon J, Coombs RW. Clinical performance of the Multispot HIV-1/HIV-2 rapid test to correctly differentiate HIV-2 from HIV-1 infection in screening algorithms using third and fourth generation assays and to identify cross reactivity with the HIV-1 Western Blot. J Clin Virol. 2013;58 Suppl 1:e104-7.
[\[PubMed Abstract\]](#) -
89. Croft P, Altman DG, Deeks JJ, et al. The science of clinical practice: disease diagnosis or patient prognosis? Evidence about "what is likely to happen" should shape clinical practice. BMC Med. 2015;13:20.
[\[PubMed Abstract\]](#) -
90. Altman DG, Bland JM. Diagnostic tests 2: Predictive values. BMJ. 1994;309:102.
[\[PubMed Abstract\]](#) -
91. Altman DG, Bland JM. Diagnostic tests. 1: Sensitivity and specificity. BMJ. 1994;308:1552.
[\[PubMed Abstract\]](#) -
92. Manak MM, Jagodzinski LL, Shutt A, et al. Decreased Seroreactivity in Individuals Initiating Antiretroviral Therapy during Acute HIV Infection. J Clin Microbiol. 2019;57:pii: e00757-19.
[\[PubMed Abstract\]](#) -
93. Kassutto S, Johnston MN, Rosenberg ES. Incomplete HIV type 1 antibody evolution and seroreversion in acutely infected individuals treated with early antiretroviral therapy. Clin Infect Dis. 2005;40:868-73.
[\[PubMed Abstract\]](#) -
94. Hare CB, Pappalardo BL, Busch MP, et al. Seroreversion in subjects receiving antiretroviral therapy

- during acute/early HIV infection. Clin Infect Dis. 2006;42:700-8.
[\[PubMed Abstract\]](#) -
95. de Souza MS, Pinyakorn S, Akapirat S, et al. Initiation of Antiretroviral Therapy During Acute HIV-1 Infection Leads to a High Rate of Nonreactive HIV Serology. Clin Infect Dis. 2016;63:555-61.
[\[PubMed Abstract\]](#) -
96. Sullivan PS, Schable C, Koch W, et al. Persistently negative HIV-1 antibody enzyme immunoassay screening results for patients with HIV-1 infection and AIDS: serologic, clinical, and virologic results. Seronegative AIDS Clinical Study Group. AIDS. 1999;13:89-96.
[\[PubMed Abstract\]](#) -
97. Spivak AM, Brennan TP, O'Connell KA, et al. A case of seronegative HIV-1 infection. J Infect Dis. 2010;201:341-5.
[\[PubMed Abstract\]](#) -
98. Spivak AM, Sydnor ER, Blankson JN, Gallant JE. Seronegative HIV-1 infection: a review of the literature. AIDS. 2010;24:1407-14.
[\[PubMed Abstract\]](#) -
99. Ellenberger DL, Sullivan PS, Dorn J, et al. Viral and immunologic examination of human immunodeficiency virus type 1-infected, persistently seronegative persons. J Infect Dis. 1999;180:1033-42.
[\[PubMed Abstract\]](#) -
100. Donnell D, Ramos E, Celum C, et al. The effect of oral preexposure prophylaxis on the progression of HIV-1 seroconversion. AIDS. 2017;31:2007-16.
[\[PubMed Abstract\]](#) -
101. Smith DK, Switzer WM, Peters P, et al. A Strategy for PrEP Clinicians to Manage Ambiguous HIV Test Results During Follow-up Visits. Open Forum Infect Dis. 2018;5:ofy180.
[\[PubMed Abstract\]](#) -
102. Padeh YC, Rubinstein A, Shliozberg J. Common variable immunodeficiency and testing for HIV-1. N Engl J Med. 2005;353:1074-5.
[\[PubMed Abstract\]](#) -
103. Jurriaans S, Sankatsing SU, Prins JM, et al. HIV-1 seroreversion in an HIV-1-seropositive patient treated during acute infection with highly active antiretroviral therapy and mycophenolate mofetil. AIDS. 2004;18:1607-8.
[\[PubMed Abstract\]](#) -
104. Roy MJ, Damato JJ, Burke DS. Absence of true seroreversion of HIV-1 antibody in seroreactive individuals. JAMA. 1993;269:2876-9.
[\[PubMed Abstract\]](#) -
105. Klarkowski D, O'Brien DP, Shanks L, Singh KP. Causes of false-positive HIV rapid diagnostic test results. Expert Rev Anti Infect Ther. 2014;12:49-62.
[\[PubMed Abstract\]](#) -
106. Alfie LG, Longueira YS, Pippo M, et al. Increased risk of false-positive HIV ELISA results after COVID-19. AIDS. 2023;37:947-50.
[\[PubMed Abstract\]](#) -

107. Theppote AS, Carmack AE, Riedel DJ. False positive HIV testing after T-cell receptor therapy. *AIDS*. 2020;34:1103-1105.
[\[PubMed Abstract\]](#) -
108. Cornett JK, Kirn TJ. Laboratory diagnosis of HIV in adults: a review of current methods. *Clin Infect Dis*. 2013;57:712-8.
[\[PubMed Abstract\]](#) -
109. Cohen MS, Gay CL, Busch MP, Hecht FM. The detection of acute HIV infection. *J Infect Dis*. 2010;202 Suppl 2:S270-7.
[\[PubMed Abstract\]](#) -
110. Centers for Disease Control and Prevention (CDC). Detection of acute HIV infection in two evaluations of a new HIV diagnostic testing algorithm - United States, 2011-2013. *MMWR Morb Mortal Wkly Rep*. 2013;62:489-94.
[\[PubMed Abstract\]](#) -
111. Masciotra S, Smith AJ, Youngpairoj AS, et al. Evaluation of the CDC proposed laboratory HIV testing algorithm among men who have sex with men (MSM) from five US metropolitan statistical areas using specimens collected in 2011. *J Clin Virol*. 2013;58 Suppl 1:e8-e12.
[\[PubMed Abstract\]](#) -
112. Schacker T, Collier AC, Hughes J, Shea T, Corey L. Clinical and epidemiologic features of primary HIV infection. *Ann Intern Med*. 1996;125:257-64.
[\[PubMed Abstract\]](#) -
113. Daar ES, Pilcher CD, Hecht FM. Clinical presentation and diagnosis of primary HIV-1 infection. *Curr Opin HIV AIDS*. 2008;3:10-5.
[\[PubMed Abstract\]](#) -
114. Daar ES, Little S, Pitt J, et al. Diagnosis of primary HIV-1 infection. Los Angeles County Primary HIV Infection Recruitment Network. *Ann Intern Med*. 2001;134:25-9.
[\[PubMed Abstract\]](#) -
115. Sivay MV, Li M, Piwowar-Manning E, et al. Characterization of HIV Seroconverters in a TDF/FTC PrEP Study: HPTN 067/ADAPT. *J Acquir Immune Defic Syndr*. 2017;75:271-9.
[\[PubMed Abstract\]](#) -
116. Curlin ME, Gvetadze R, Leelawiwat W, et al. Analysis of False-Negative Human Immunodeficiency Virus Rapid Tests Performed on Oral Fluid in 3 International Clinical Research Studies. *Clin Infect Dis*. 2017;64:1663-9.
[\[PubMed Abstract\]](#) -
117. Landovitz RJ, Donnell D, Clement ME, et al. Cabotegravir for HIV prevention in cisgender men and transgender women. *N Engl J Med*. 2021;385:595-608.
[\[PubMed Abstract\]](#) -
118. Peruski AH, Wesolowski LG, Delaney KP, et al. Trends in HIV-2 Diagnoses and Use of the HIV-1/HIV-2 Differentiation Test - United States, 2010-2017. *MMWR Morb Mortal Wkly Rep*. 2020;69:63-6.
[\[PubMed Abstract\]](#) -
119. Panel on Antiretroviral Guidelines for Adults and Adolescents. Guidelines for the use of antiretroviral agents in adults and adolescents with HIV. Department of Health and Human Services. Considerations for antiretroviral use in special patient populations: HIV-2 infection. December 6, 2023.

[\[HIV.gov\]](#) -

120. Centers for Disease Control and Prevention. HIV-2 Infection Surveillance--United States, 1987-2009. MMWR Morb Mortal Wkly Rep. 2011;60:985-8.
[\[PubMed Abstract\]](#) -
121. O'Brien TR, George JR, Epstein JS, Holmberg SD, Schochetman G. Testing for antibodies to human immunodeficiency virus type 2 in the United States. MMWR Recomm Rep. 1992;41:1-9.
[\[PubMed Abstract\]](#) -
122. Bissessor M, Bradshaw CS, Fairley CK, Chen MY, Chow EP. Provision of HIV test results by telephone is both safe and efficient for men who have sex with men. Int J STD AIDS. 2017;28:39-44.
[\[PubMed Abstract\]](#) -
123. McKinstry LA, Goldbaum GM, Meischke HW. Telephone notification of HIV test results: impact in King County, Washington. Sex Transm Dis. 2007;34:796-800.
[\[PubMed Abstract\]](#) -
124. Thornton AC, Delpech V, Kall MM, Nardone A. HIV testing in community settings in resource-rich countries: a systematic review of the evidence. HIV Med. 2012;13:416-26.
[\[PubMed Abstract\]](#) -
125. Rodger AJ, McCabe L, Phillips AN, et al. Free HIV self-test for identification and linkage to care of previously undetected HIV infection in men who have sex with men in England and Wales (SELPHI): an open-label, internet-based, randomised controlled trial. Lancet HIV. 2022;9:e838-e847.
[\[PubMed Abstract\]](#) -
126. Witzel TC, Eshun-Wilson I, Jamil MS, et al. Comparing the effects of HIV self-testing to standard HIV testing for key populations: a systematic review and meta-analysis. BMC Med. 2020;18:381.
[\[PubMed Abstract\]](#) -
127. Branson BM, Handsfield HH, Lampe MA, et al. Revised recommendations for HIV testing of adults, adolescents, and pregnant women in health-care settings. MMWR Recomm Rep. 2006;55:1-17.
[\[PubMed Abstract\]](#) -
128. Joseph HA, Fasula AM, Morgan RL, et al. "The anticipation alone could kill you": past and potential clients' perspectives on HIV testing in non-health care settings. AIDS Educ Prev. 2011;23:577-94.
[\[PubMed Abstract\]](#) -

References

- Bowles KE, Clark HA, Tai E, et al. Implementing rapid HIV testing in outreach and community settings: results from an advancing HIV prevention demonstration project conducted in seven U.S. cities. Public Health Rep. 2008;123 Suppl 3:78-85.
[\[PubMed Abstract\]](#) -
- Burgard M, Blanche S, Jasseron C, et al. Performance of HIV-1 DNA or HIV-1 RNA tests for early diagnosis of perinatal HIV-1 infection during anti-retroviral prophylaxis. J Pediatr. 2012;160:60-6.e1.
[\[PubMed Abstract\]](#) -
- Campbell-Yesufu OT, Gandhi RT. Update on human immunodeficiency virus (HIV)-2 Infection. Clin Infect Dis. 2011;52:780-7.
[\[PubMed Abstract\]](#) -

- CDC Testing 2013. Centers for Disease Control and Prevention. CDC-Funded HIV Testing: United States, Puerto Rico and the U.S. Virgin Islands, 2013. [\[CDC\]](#) -
- Centers for Disease Control and Prevention (CDC). False-positive oral fluid rapid HIV tests--New York City, 2005-2008. MMWR Morb Mortal Wkly Rep. 2008;57:660-5. [\[MMWR\]](#) -
- Centers for Disease Control and Prevention. Diagnoses of HIV infection in the United States and dependent areas, 2021. HIV Surveillance Report, 2021; vol. 34. Published May 2023. [\[CDC\]](#) -
- Centers for Disease Control and Prevention. Estimated HIV Incidence and Prevalence in the United States, 2015-2019. HIV Surveillance Supplemental Report. 2021;26(No. 1):1-81. Published May 2021. [\[CDC\]](#) -
- Chappel RJ, Wilson KM, Dax EM. Immunoassays for the diagnosis of HIV: meeting future needs by enhancing the quality of testing. Future Microbiol. 2009;4:963-82. [\[PubMed Abstract\]](#) -
- D'Angelo AB, Morrison CA, Lopez-Rios J, et al. Experiences Receiving HIV-Positive Results by Phone: Acceptability and Implications for Clinical and Behavioral Research. AIDS Behav. 2021;25:709-20. [\[PubMed Abstract\]](#) -
- De Clercq J, Rutsaert S, De Scheerder MA, Verhofstede C, Callens S, Vandekerckhove L. Benefits of antiretroviral therapy initiation during acute HIV infection. Acta Clin Belg. 2020:1-9. [\[PubMed Abstract\]](#) -
- Eshleman SH, Khaki L, Laeyendecker O, et al. Detection of individuals with acute HIV-1 infection using the ARCHITECT HIV Ag/Ab Combo assay. J Acquir Immune Defic Syndr. 2009;52:121-4. [\[PubMed Abstract\]](#) -
- Havens PL, Mofenson LM, American Academy of Pediatrics Committee on Pediatric AIDS. Evaluation and management of the infant exposed to HIV-1 in the United States. Pediatrics. 2009;123:175-87. [\[PubMed Abstract\]](#) -
- Hutchinson AB, Ethridge SF, Wesolowski LG, et al. Costs and outcomes of laboratory diagnostic algorithms for the detection of HIV. J Clin Virol. 2013;58 Suppl 1:e2-7. [\[PubMed Abstract\]](#) -
- Kwon JA, Yoon SY, Lee CK, et al. Performance evaluation of three automated human immunodeficiency virus antigen-antibody combination immunoassays. J Virol Methods. 2006;133:20-6. [\[PubMed Abstract\]](#) -
- Linley L, Ethridge SF, Oraka E, et al. Evaluation of supplemental testing with the Multispot HIV-1/HIV-2 Rapid Test and APTIMA HIV-1 RNA Qualitative Assay to resolve specimens with indeterminate or negative HIV-1 Western blots. J Clin Virol. 2013;58 Suppl 1:e108-12. [\[PubMed Abstract\]](#) -
- Ly TD, Laperche S, Brennan C, et al. Evaluation of the sensitivity and specificity of six HIV combined p24 antigen and antibody assays. J Virol Methods. 2004;122:185-94. [\[PubMed Abstract\]](#) -

- Mitchell EO, Stewart G, Bajzik O, Ferret M, Bentsen C, Shriver MK. Performance comparison of the 4th generation Bio-Rad Laboratories GS HIV Combo Ag/Ab EIA on the EVOLIS™ automated system versus Abbott ARCHITECT HIV Ag/Ab Combo, Ortho Anti-HIV 1+2 EIA on Vitros ECI and Siemens HIV-1/O/2 enhanced on Advia Centaur. J Clin Virol. 2013;58 Suppl 1:e79-84.
[PubMed Abstract] -
- Nasrullah M, Wesolowski LG, Meyer WA 3rd, et al. Performance of a fourth-generation HIV screening assay and an alternative HIV diagnostic testing algorithm. AIDS. 2013;27:731-7.
[PubMed Abstract] -
- Nyamweya S, Hegedus A, Jaye A, Rowland-Jones S, Flanagan KL, Macallan DC. Comparing HIV-1 and HIV-2 infection: Lessons for viral immunopathogenesis. Rev Med Virol. 2013;23:221-40.
[PubMed Abstract] -
- Pandori MW, Westheimer E, Gay C, et al. The Multispot rapid HIV-1/HIV-2 differentiation assay is comparable with the Western blot and an immunofluorescence assay at confirming HIV infection in a prospective study in three regions of the United States. J Clin Virol. 2013;58 Suppl 1:e92-6.
[PubMed Abstract] -
- Panel on Antiretroviral Guidelines for Adults and Adolescents. Guidelines for the use of antiretroviral agents in adults and adolescents with HIV. Department of Health and Human Services. Initiation of antiretroviral therapy. December 18, 2019.
[HIV.gov] -
- Panel on Antiretroviral Therapy and Medical Management of Children Living with HIV. Guidelines for the use of antiretroviral agents in pediatric HIV infection. Diagnosis of HIV infection in infants and children. January 31, 2023.
[HIV.gov] -
- Panneer N, Lontok E, Branson BM, et al. HIV and hepatitis C virus infection in the United States: whom and how to test. Clin Infect Dis. 2014;59:875-82.
[PubMed Abstract] -
- Patel P, Mackellar D, Simmons P, et al. Detecting acute human immunodeficiency virus infection using 3 different screening immunoassays and nucleic acid amplification testing for human immunodeficiency virus RNA, 2006-2008. Arch Intern Med. 2010;170:66-74.
[PubMed Abstract] -
- Ramos EM, Harb S, Dragavon J, Swenson P, Stekler JD, Coombs RW. Performance of an alternative HIV diagnostic algorithm using the ARCHITECT HIV Ag/Ab Combo assay and potential utility of sample-to-cutoff ratio to discriminate primary from established infection. J Clin Virol. 2013;58 Suppl 1:e38-43.
[PubMed Abstract] -
- Rosenberg NE, Pilcher CD, Busch MP, Cohen MS. How can we better identify early HIV infections? Curr Opin HIV AIDS. 2015;10:61-8.
[PubMed Abstract] -
- Schappert J, Wians FH Jr, Schiff E, et al. Multicenter evaluation of the Bayer ADVIA Centaur HIV 1/O/2 enhanced (EHIV) assay. Clin Chim Acta. 2006;372:158-66.
[PubMed Abstract] -
- Smith MK, Rutstein SE, Powers KA, et al. The detection and management of early HIV infection: a clinical and public health emergency. J Acquir Immune Defic Syndr. 2013;63 Suppl 2:S187-99.
[PubMed Abstract] -

- Spielberg F, Levine RO, Weaver M. Self-testing for HIV: a new option for HIV prevention? *Lancet Infect Dis.* 2004;4:640-6.
[\[PubMed Abstract\]](#) -
- Torian LV, Forgione LA, Punsalang AE, Pirillo RE, Oleszko WR. Comparison of Multispot EIA with Western blot for confirmatory serodiagnosis of HIV. *J Clin Virol.* 2011;52 Suppl 1:S41-4.
[\[PubMed Abstract\]](#) -
- Tudor Car L, Gentry S, van-Velthoven MH, Car J. Telephone communication of HIV testing results for improving knowledge of HIV infection status. *Cochrane Database Syst Rev.* 2013;1:CD009192.
[\[PubMed Abstract\]](#) -
- U.S. Food and Drug Administration. ADVIA Centaur HIV 1/O/2 Enhanced ReadyPack Reagents.
[\[U.S. FDA\]](#) -
- U.S. Food and Drug Administration. Avioq HIV-1 Microelisa System.
[\[U.S. FDA\]](#) -
- U.S. Food and Drug Administration. Genetic Systems HIV-1/HIV-2 Plus O EIA.
[\[U.S. FDA\]](#) -
- U.S. Food and Drug Administration. Home Access HIV-1 Test System.
[\[U.S. FDA\]](#) -
- U.S. Food and Drug Administration. VITROS Immunodiagnosics Products Anti-HIV 1+2 Calibrator VITROS Immunodiagnosics Products Anti-HIV 1+2 Reagent Pack.
[\[U.S. FDA\]](#) -
- Udeagu CC, Bocour A, Shepard CW. For partner services, do we need "face-time," or FaceTime?: Trends in relative proportion of in-person notifications and HIV testing after introduction of a telephone option for HIV exposure notification by public health. *Sex Transm Dis.* 2014;41:671-3.
[\[PubMed Abstract\]](#) -

Figures

Figure 1 HIV Care Continuum

Source: Adapted from HRSA. HIV Care Continuum

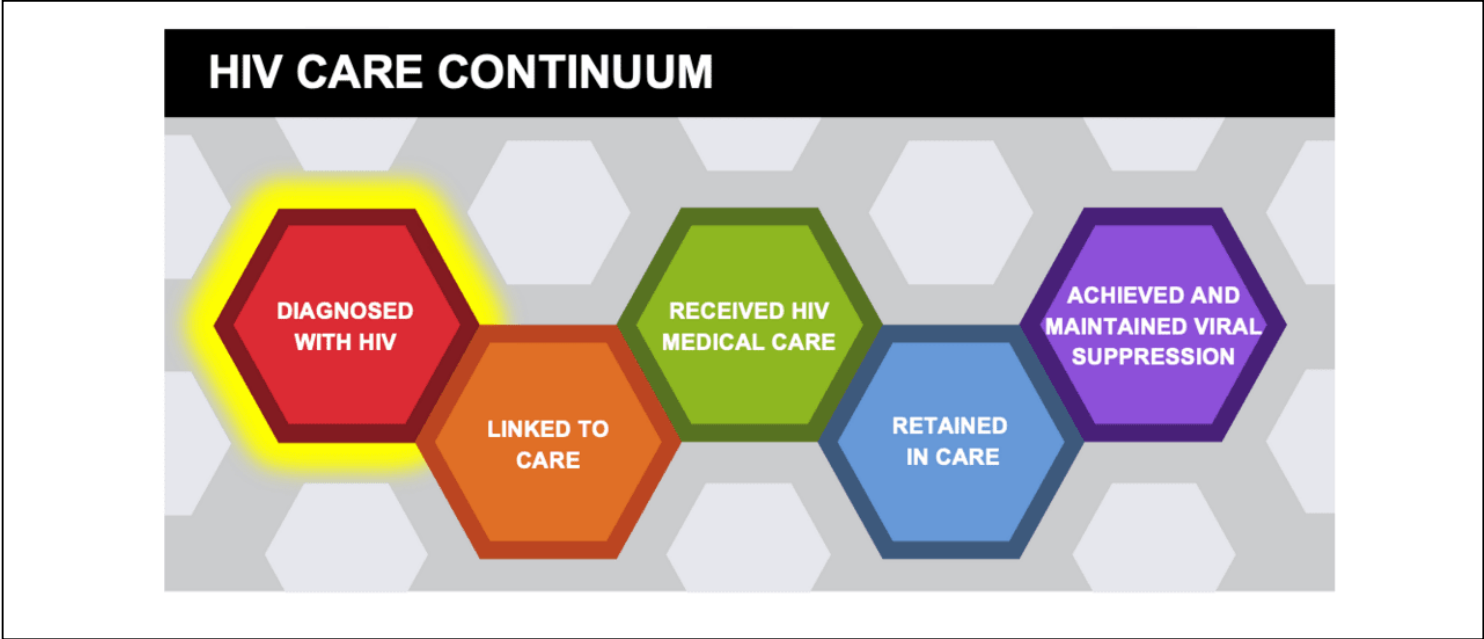


Figure 2 (Image Series) - Fiebig Classification for Early HIV-1 Infection (Image Series) - Figure 2 (Image Series) - Fiebig Classification for Early HIV-1 Infection

Image 2A: Duration of Stages

Source: Fiebig EW, Wright DJ, Rawal BD, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. AIDS. 2003;17:1871-9.

Fiebig Laboratory Stages of Early HIV-1 Infection					
Stage	Duration	HIV RNA	p24 Ag	*EIA	Western blot
Eclipse	11.0 days	(-)	(-)	(-)	(-)
I	5.0 days	(+)	(-)	(-)	(-)
II	5.3 days	(+)	(+)	(-)	(-)
III	3.2 days	(+)	(+)	(+)	(-)
IV	5.6 days	(+)	(+)	(+/-)	#Indeterminate pattern
V	69.5 days	(+)	(+/-)	(+)	Reactive, but absence of p31 (<i>pol</i>)
VI	Open-ended	(+)	(+/-)	(+)	Reactive, including p31 (<i>pol</i>)

* EIA = enzyme immunoassay (refers to IgM-sensitive 3rd generation assay)
 # Indeterminate Western blot: presence of HIV-1 specific bands that fail to meet criteria established by US FDA for positive HIV (reactivity to two of the following three bands: p24, gp41, gp120/160)

Figure 2 (Image Series) - Fiebig Classification for Early HIV-1 Infection
Image 2B: Graphic of Stages

Source: Fiebig EW, Wright DJ, Rawal BD, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. AIDS. 2003;17:1871-9.

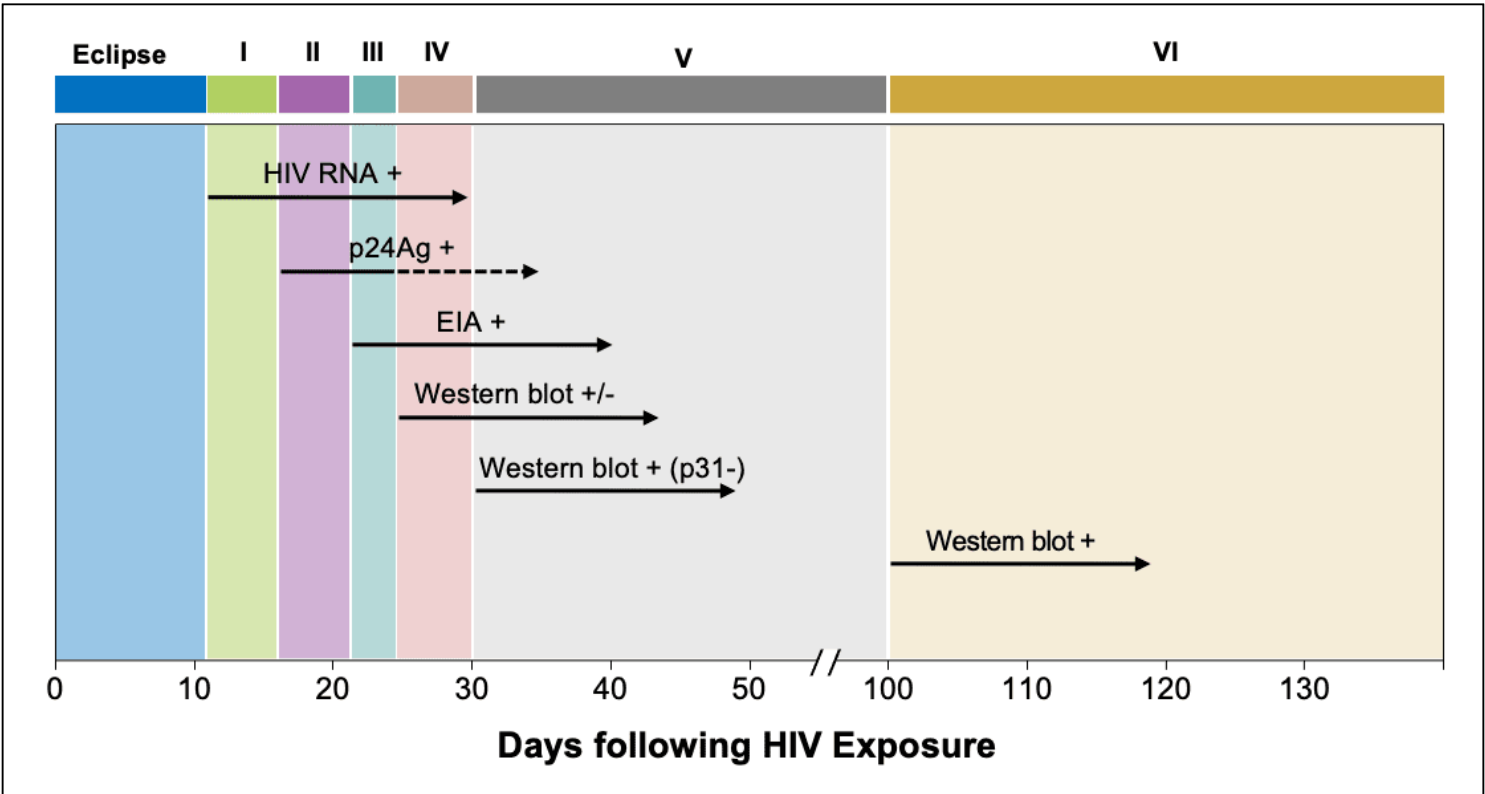


Figure 3 HIV Eclipse Phase

Illustration: David H. Spach, MD

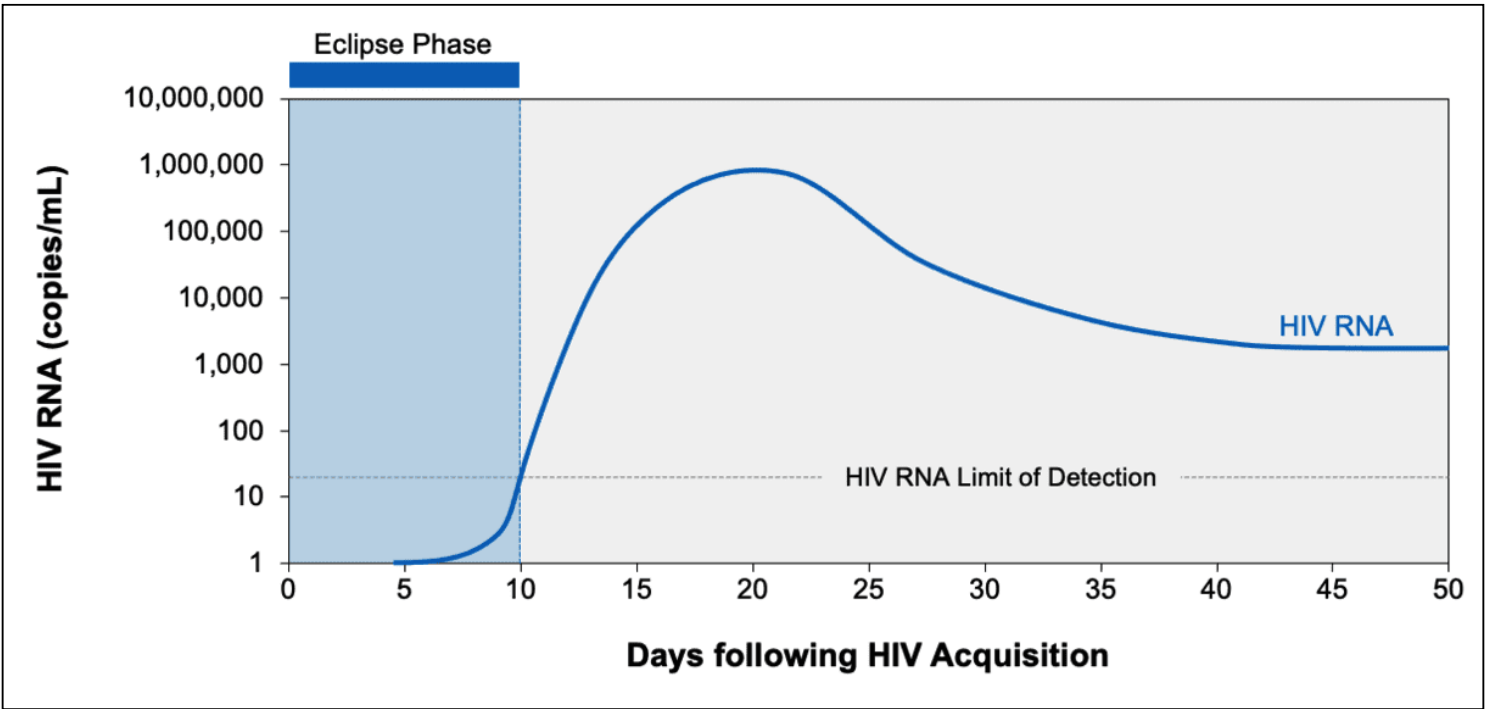


Figure 4 HIV Seroconversion Window

Illustration: David H. Spach, MD

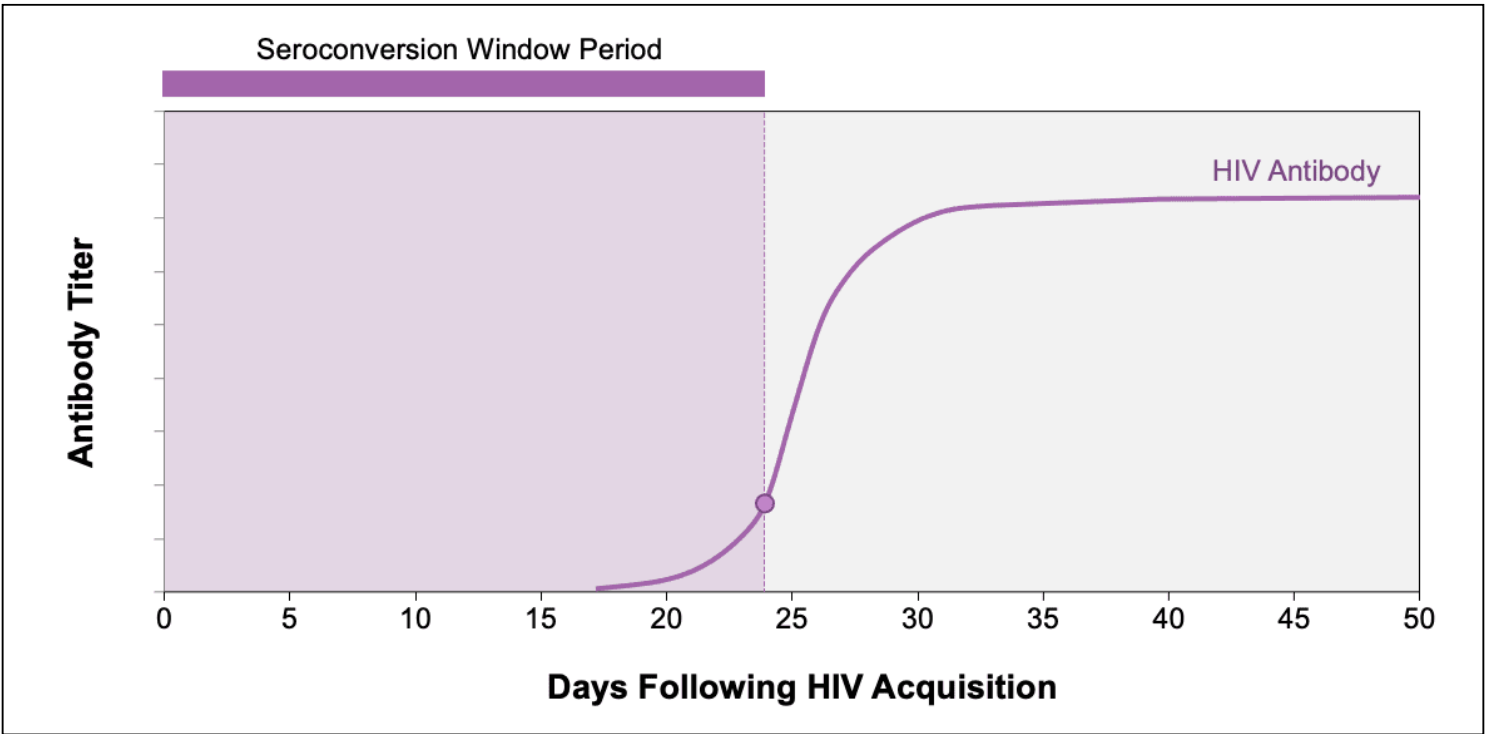


Figure 5 Timing of Positivity for HIV Diagnostic Tests

This graphic shows estimates for the mean number of days for HIV diagnostic tests to become positive after acquisition of HIV.

Abbreviation: POC = point-of-care

Source: modified from Centers for Disease Control and Prevention and Association of Public Health Laboratories. Laboratory Testing for the Diagnosis of HIV Infection: Updated Recommendations. Published June 27, 2014.

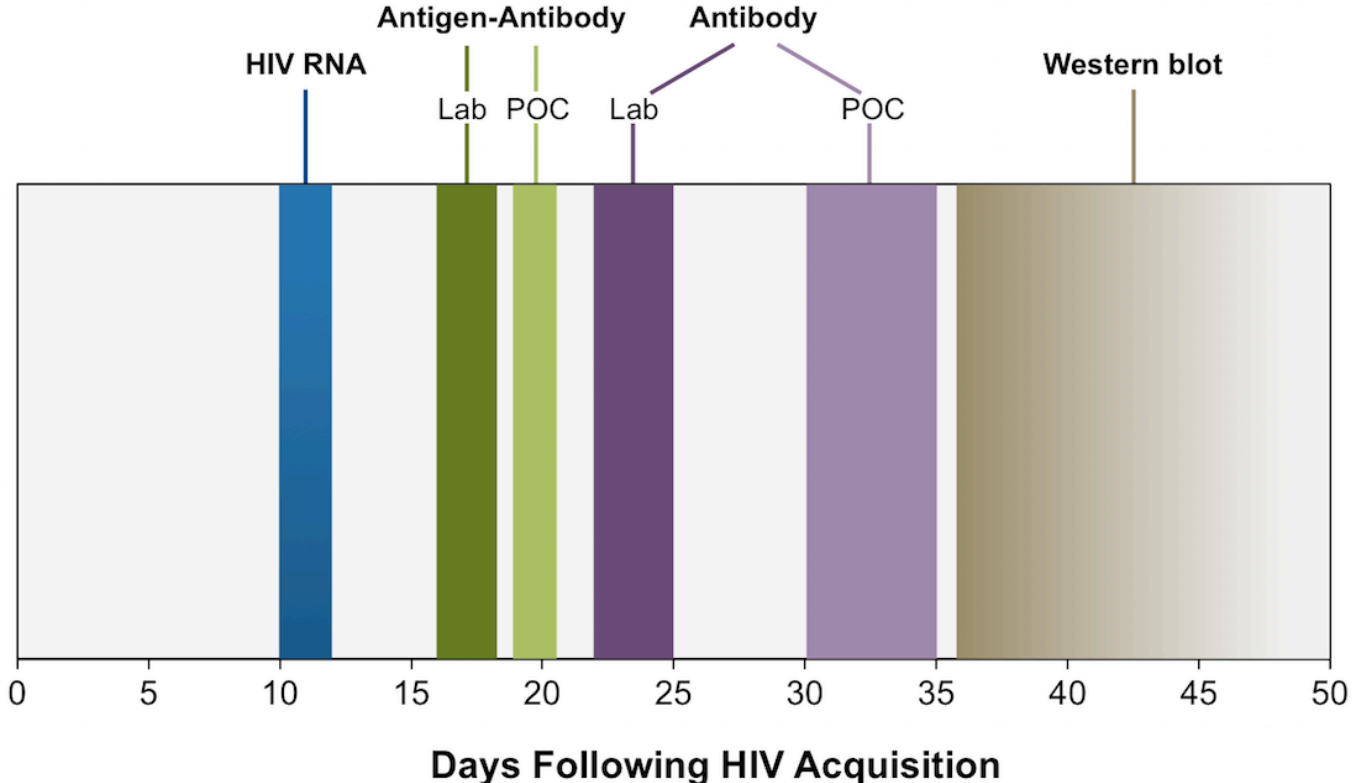


Figure 6 HIV Serologic Tests: IgG-Sensitive, IgM-Sensitive, and Antigen-Antibody

Illustration: David H. Spach, MD

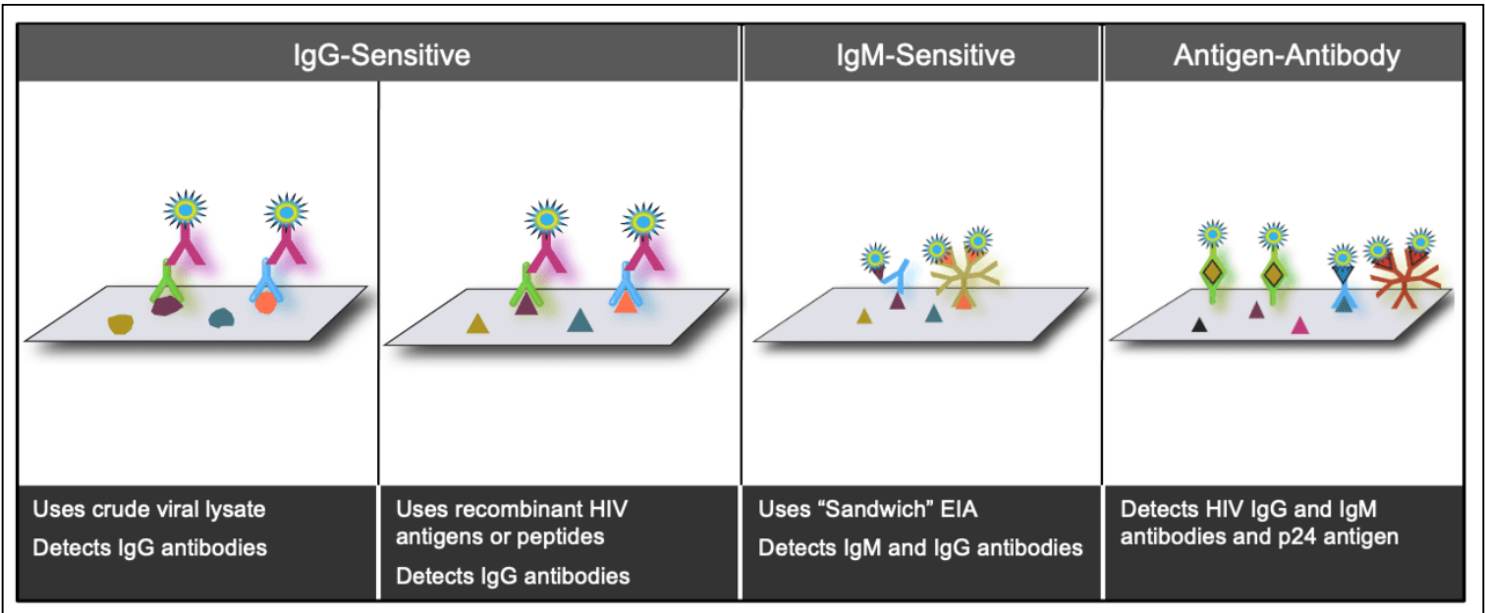


Figure 7 (Image Series) - Laboratory-Based HIV-1/2 Antigen-Antibody Immunoassays (Image Series) - Figure 7 (Image Series) - Laboratory-Based HIV-1/2 Antigen-Antibody Immunoassays Image 7A: Components of HIV-1/2 Antigen-Antibody Immunoassay

The HIV-1/2 antigen-antibody immunoassay contains components that will detect HIV-1 p24 antigen, antibodies to HIV-1, and antibodies to HIV-2. The HIV-1 and HIV-2 recombinant proteins vary from assay to assay.

Illustration by David H. Spach, MD

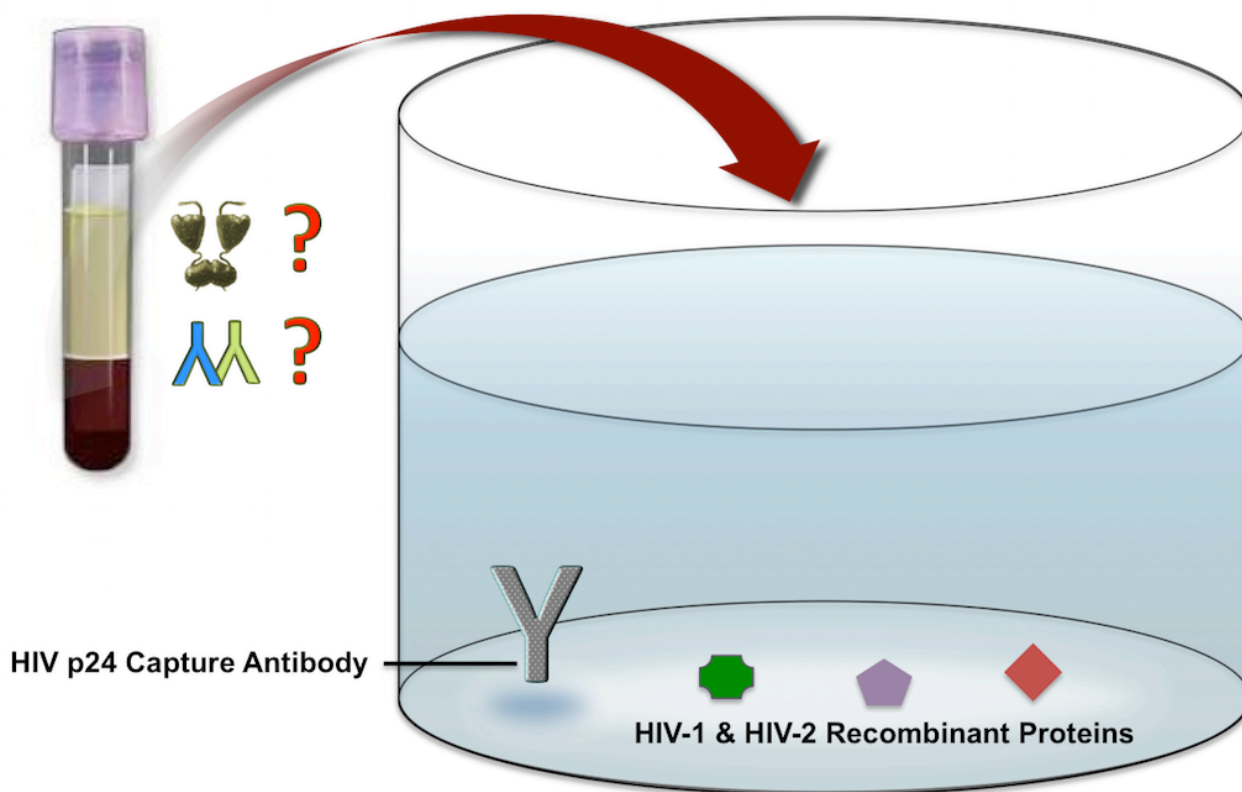


Figure 7 (Image Series) - Laboratory-Based HIV-1/2 Antigen-Antibody Immunoassays
Image 7B: Patient Sample Reacting with Components in HIV-1/2 Antigen-Antibody Immunoassay

In this example, the patient sample contains HIV-1 p24 antigen and anti-HIV antibodies that bind to the HIV-1 p24 capture antibody and the HIV recombinant proteins.

Illustration: David H. Spach, MD

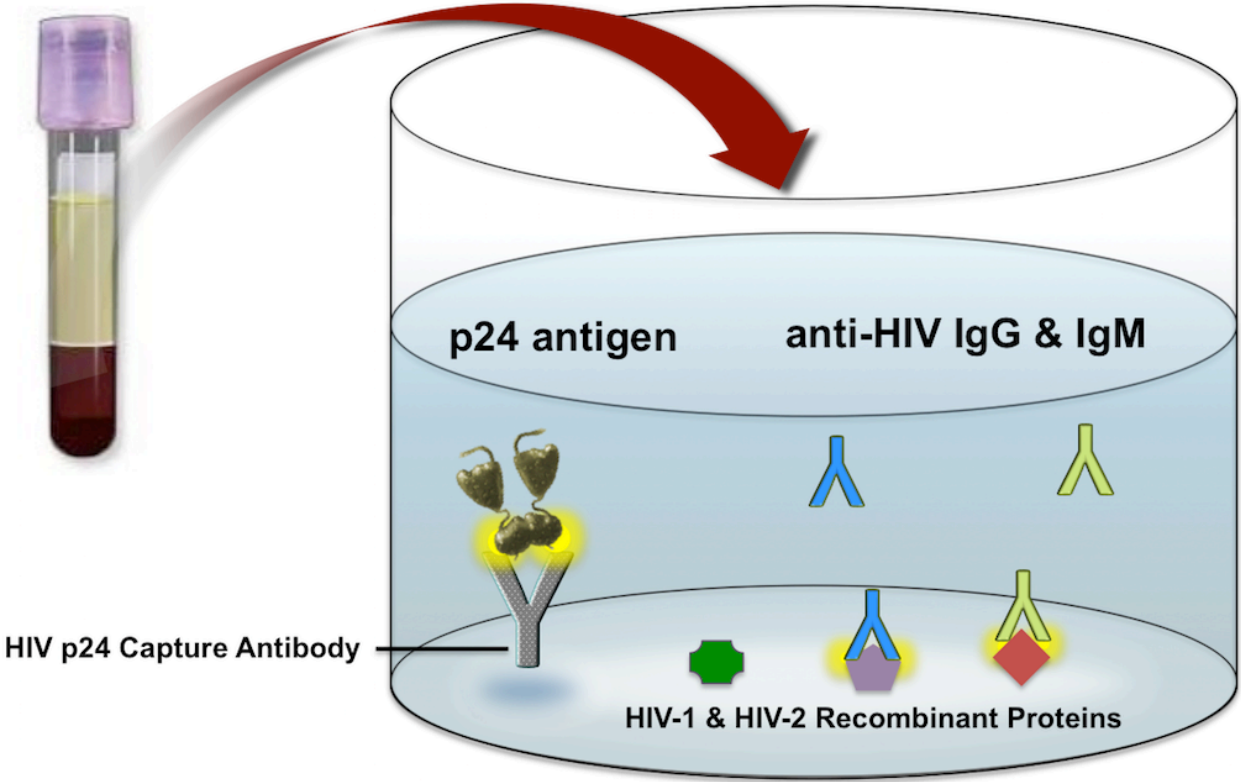


Figure 7 (Image Series) - Laboratory-Based HIV-1/2 Antigen-Antibody Immunoassays
Image 7C: Reactive HIV-1/2 Antigen-Antibody Immunoassay

The HIV-1/2 antigen-antibody immunoassay will turn positive with the presence of one or more of the following: HIV-1 p24 antigen, antibodies to HIV-1, or antibodies to HIV-2. With most of the laboratory-based assays, the positive reaction is nonspecific and thus does not differentiate HIV-1 p24 antigen, antibodies to HIV-1, or antibodies to HIV-2. In addition, most of the assays will not determine whether more than one of these components are present in a positive reaction.

Illustration: David H. Spach, MD

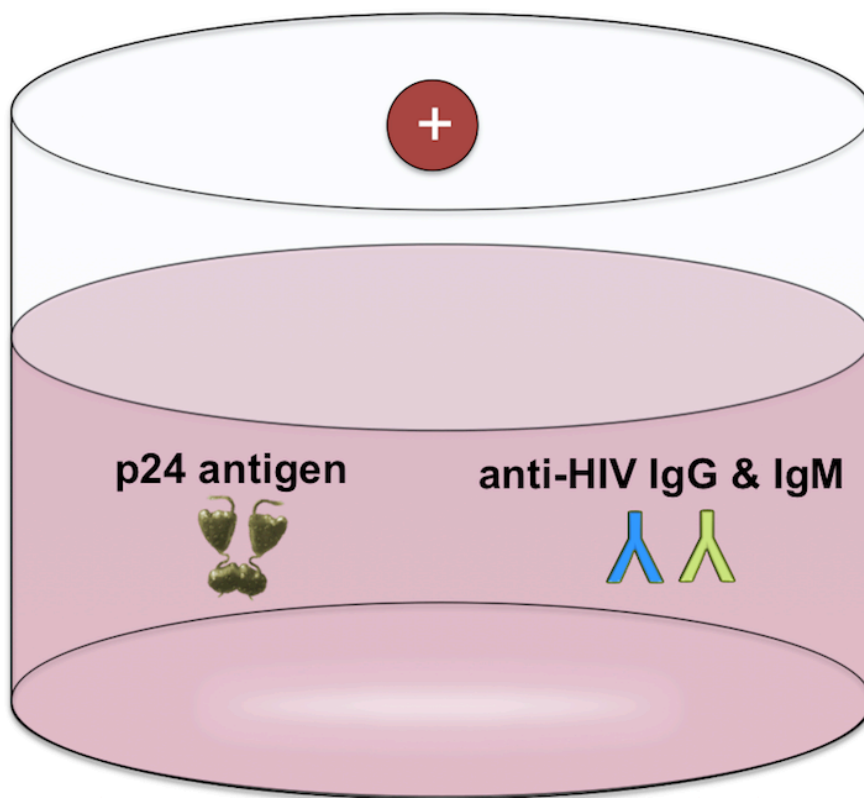


Figure 8 Timing of HIV-1/2 Antigen-Antibody Immunoassay Reactivity Following HIV Acquisition

This graphic shows the time course for test HIV-1/2 antigen-antibody immunoassay positivity after HIV acquisition: 25% at day 14, 50% at day 18, 75% at day 24, and 99% after day 44. Thus, a negative test at day 45 after an exposure virtually excludes HIV infection from that exposure.

Source: Delaney KP, Hanson DL, Masciotra S, Ethridge SF, Wesolowski L, Owen SM. Time Until Emergence of HIV Test Reactivity Following Infection With HIV-1: Implications for Interpreting Test Results and Retesting After Exposure. Clin Infect Dis. 2017;64:53-9.

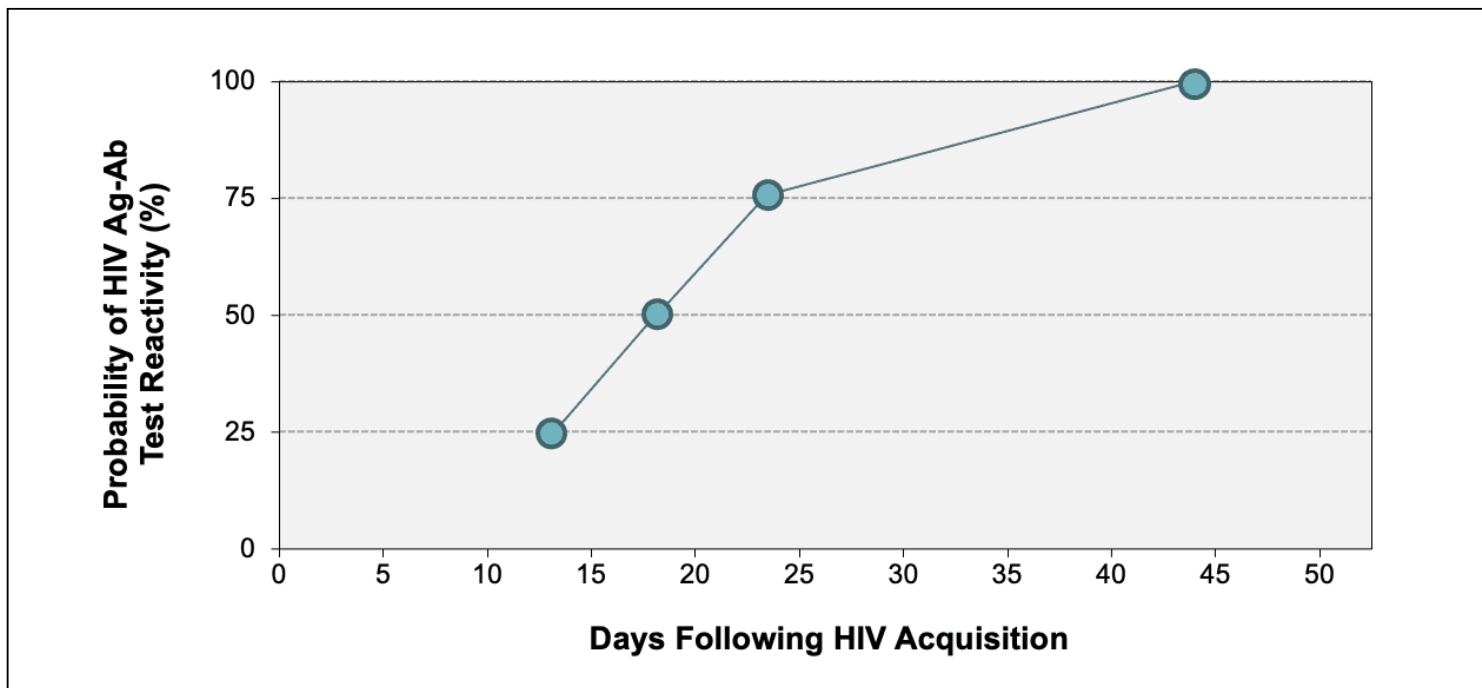


Figure 9 Geenius HIV 1/2 Supplemental Assay

Source: modified from Fernández McPhee C, Álvarez P, Prieto L, et al. HIV-1 infection using dried blood spots can be confirmed by Bio-Rad Geenius™ HIV 1/2 confirmatory assay. J Clin Virol. 2015;63:66-9.

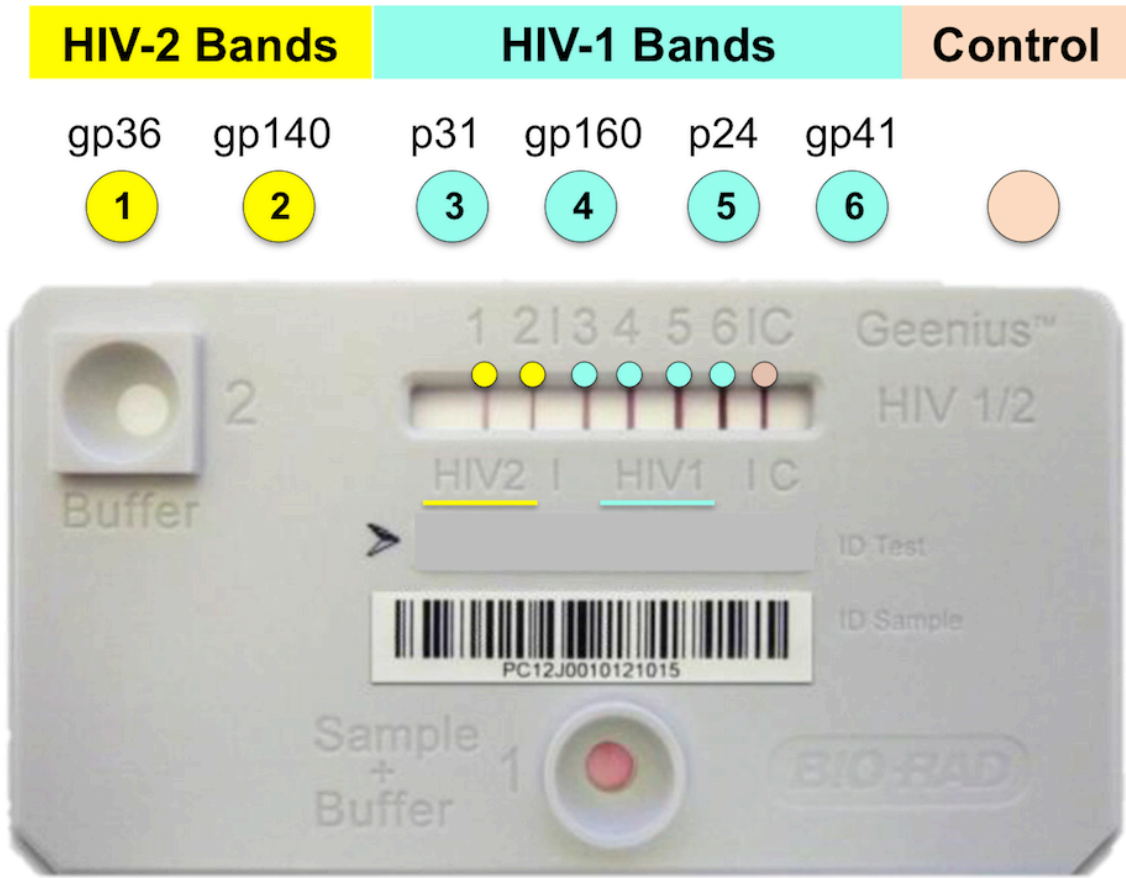


Figure 10 HIV-1 Western blot

This graphic shows the relationship of the HIV-1 genes and products with the corresponding band on the HIV-1 Western blot.

HIV-1 Gene and Product	Band on Western blot
env	
Precursor Protein	gp160
External Glycoprotein	gp120
Transmembrane Protein	gp41
pol	
Reverse Transcriptase	p66
Reverse Transcriptase	p51
Endonuclease	p31
gag	
Gag Precursor	p55
Core	p24
Matrix	p17
Nucleocapsid Precursor	p15

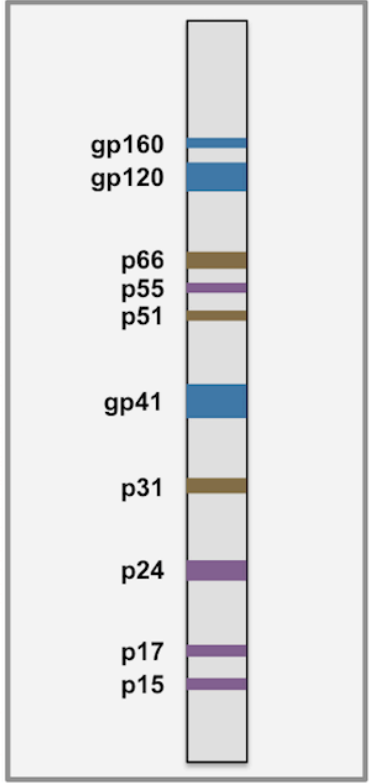


Figure 11 CDC and APHL Recommended Laboratory Testing Algorithm for the Diagnosis of HIV Infection

Source: Centers for Disease Control and Prevention and Association of Public Health Laboratories. 2018 Quick reference guide: Recommended laboratory HIV testing algorithm for serum or plasma specimens. Published January 27, 2018.

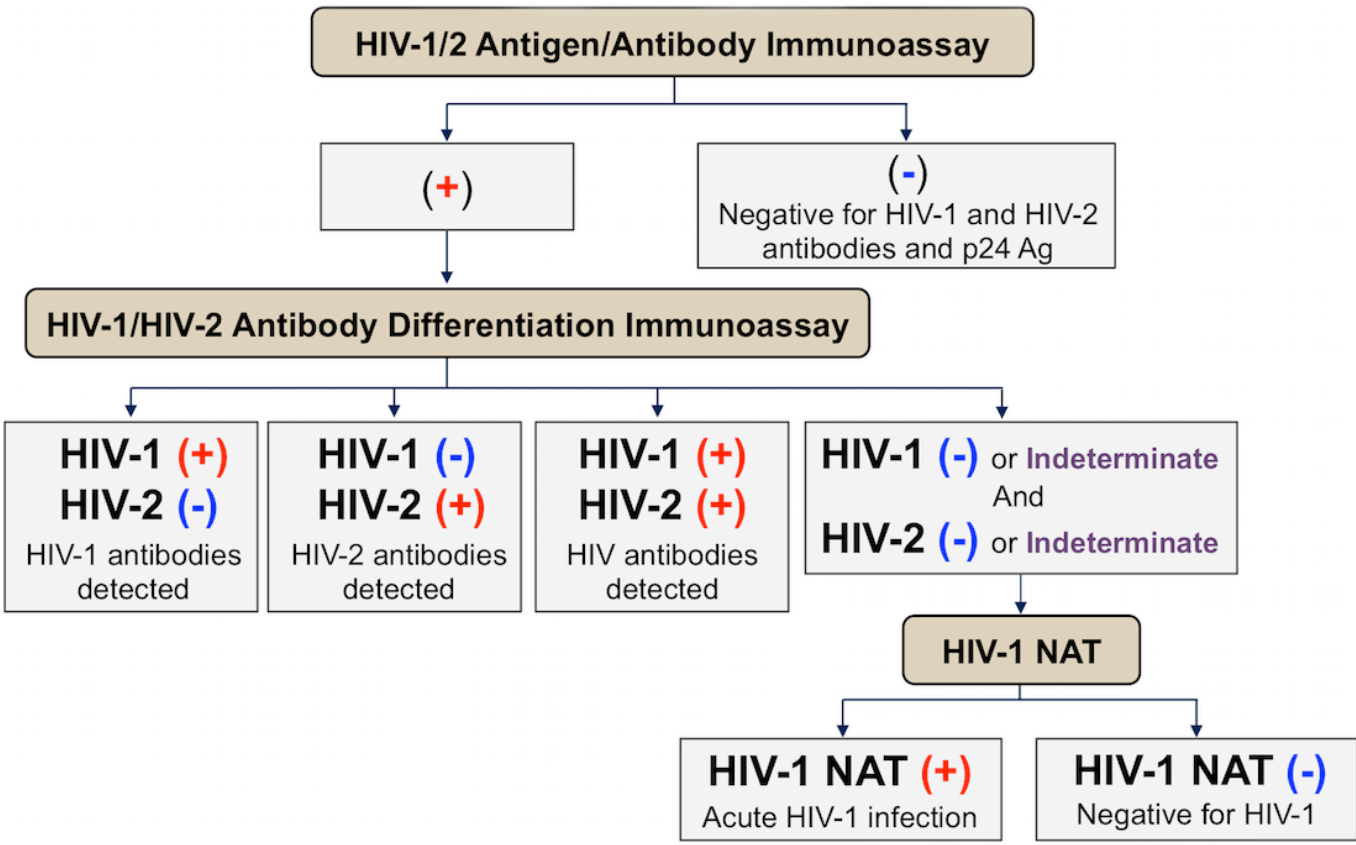


Figure 12 (Image Series) - Sensitivity of HIV Diagnostic Test (Image Series) - Figure 12 (Image Series) - Sensitivity of HIV Diagnostic Test
Image 12A: Example of Sensitivity of HIV Diagnostic Test

In this example, there are 50 persons who have HIV that are undergoing testing. Among the 50 tested, there are 49 true positives and 1 false-negative test result.

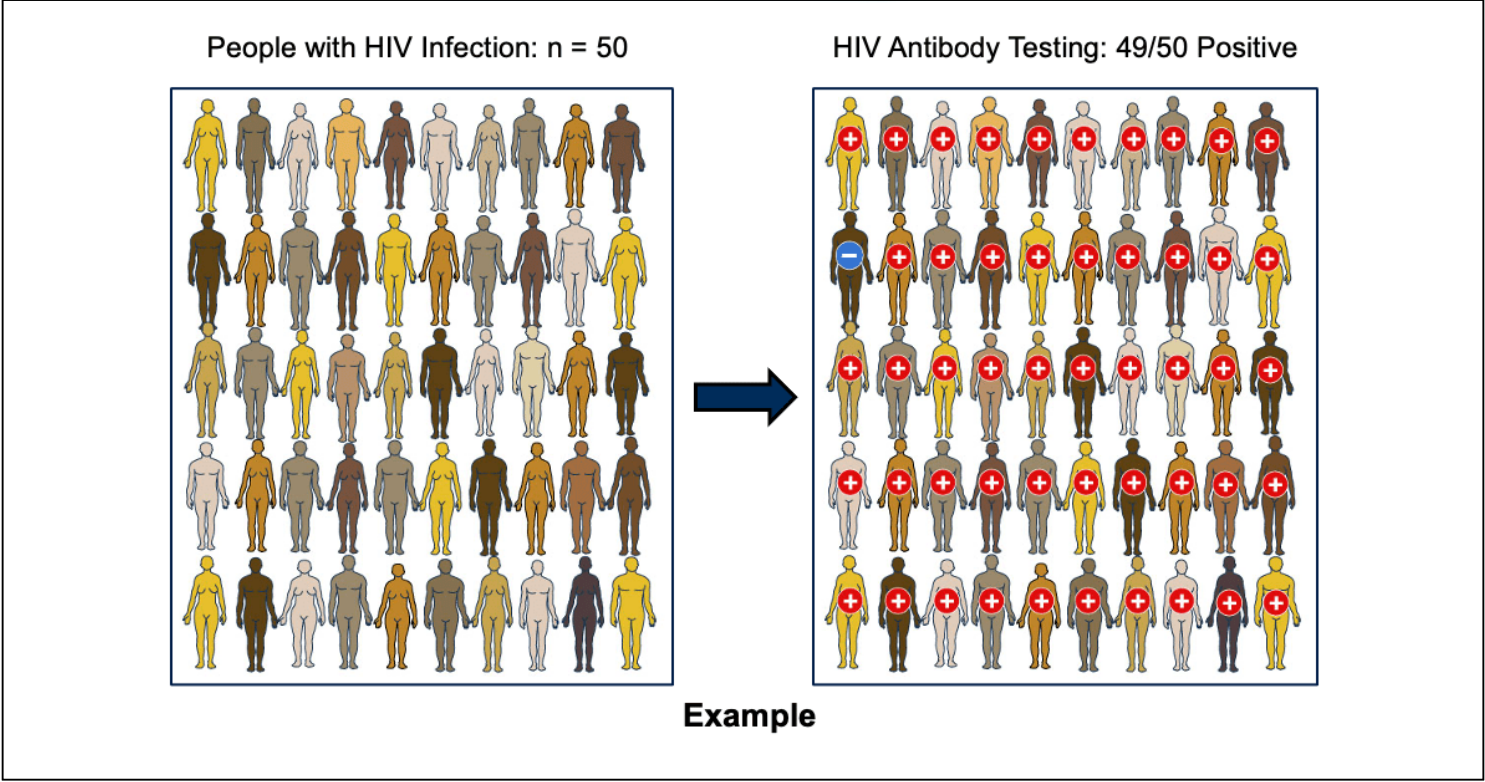


Figure 12 (Image Series) - Sensitivity of HIV Diagnostic Test
Image 12B: Sensitivity: Mathematical Expression

$$\text{Sensitivity} = \frac{\text{number of true positives}}{\text{number of true positives} + \text{number of false negatives}}$$

Figure 13 (Image Series) - Specificity of HIV Diagnostic Test (Image Series) - Figure 13 (Image Series) - Specificity of HIV Diagnostic Test
Image 13A: Example of Specificity of HIV Diagnostic Test

In this example, there are 50 persons without HIV that are undergoing HIV testing. Among the 50 tested, there are 48 true negative and 2 false-positive test results.

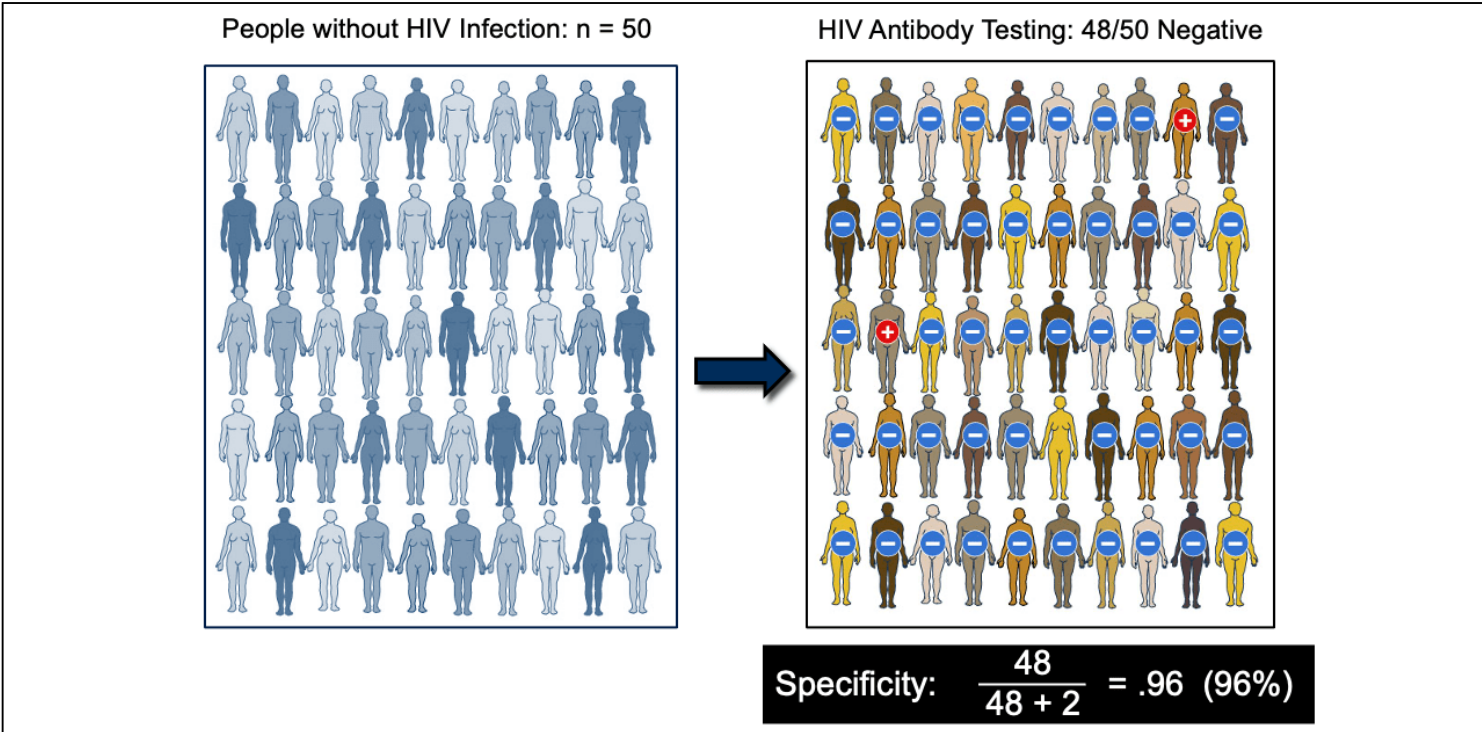


Figure 13 (Image Series) - Specificity of HIV Diagnostic Test
Image 13B: Specificity: Mathematical Expression

$$\text{Specificity} = \frac{\text{number of true negatives}}{\text{number of true negatives} + \text{number of false positives}}$$

Figure 14 (Image Series) - False-Negative HIV Diagnostic Test (Image Series) - Figure 14 (Image Series) - False-Negative HIV Diagnostic Test
Image 14A: Test Results for Persons with HIV Infection

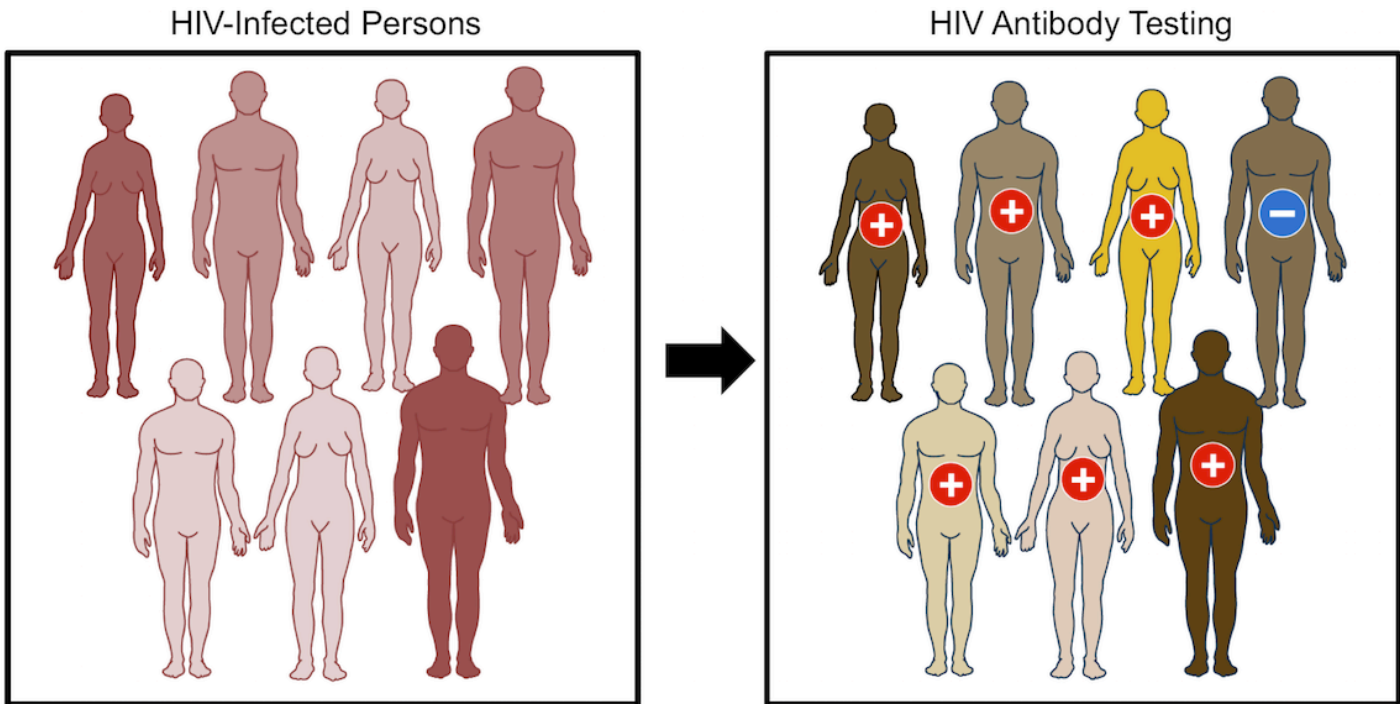


Figure 14 (Image Series) - False-Negative HIV Diagnostic Test
Image 14B: False-Negative Identified

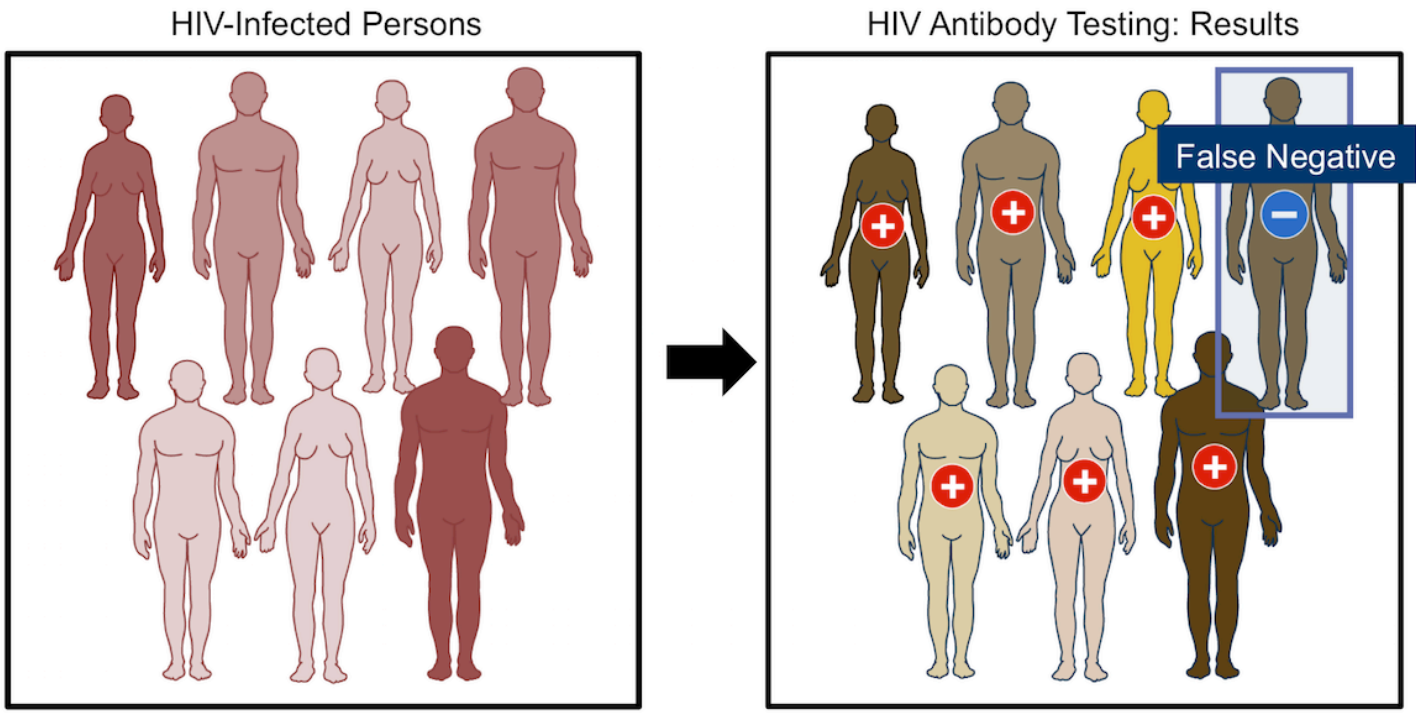


Figure 15 (Image Series) - False-Positive HIV Diagnostic Test (Image Series) - Figure 15 (Image Series) - False-Positive HIV Diagnostic Test
Image 15A: Test Results for Persons without HIV

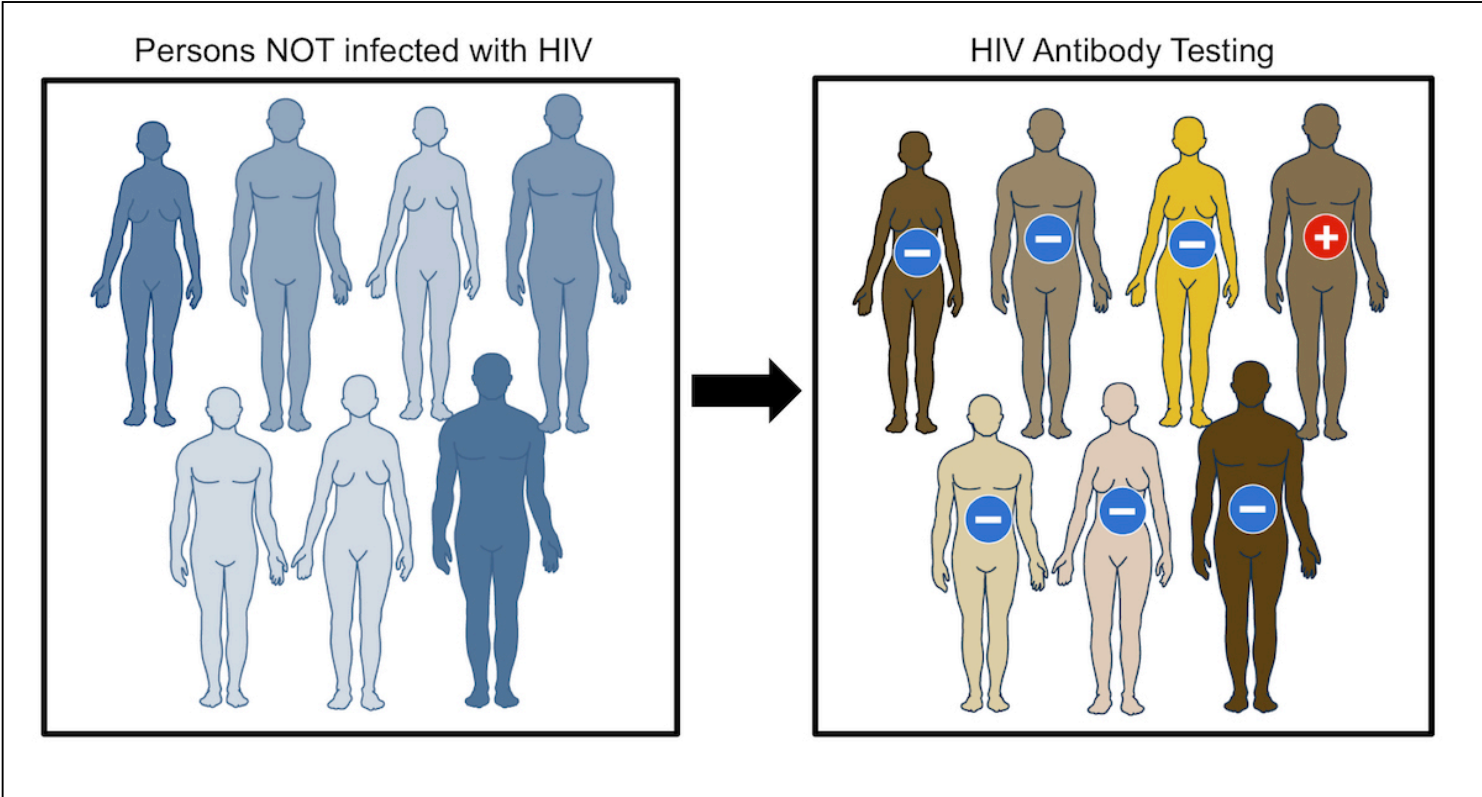


Figure 15 (Image Series) - False-Positive HIV Diagnostic Test
Image 15B: False-Positive Identified

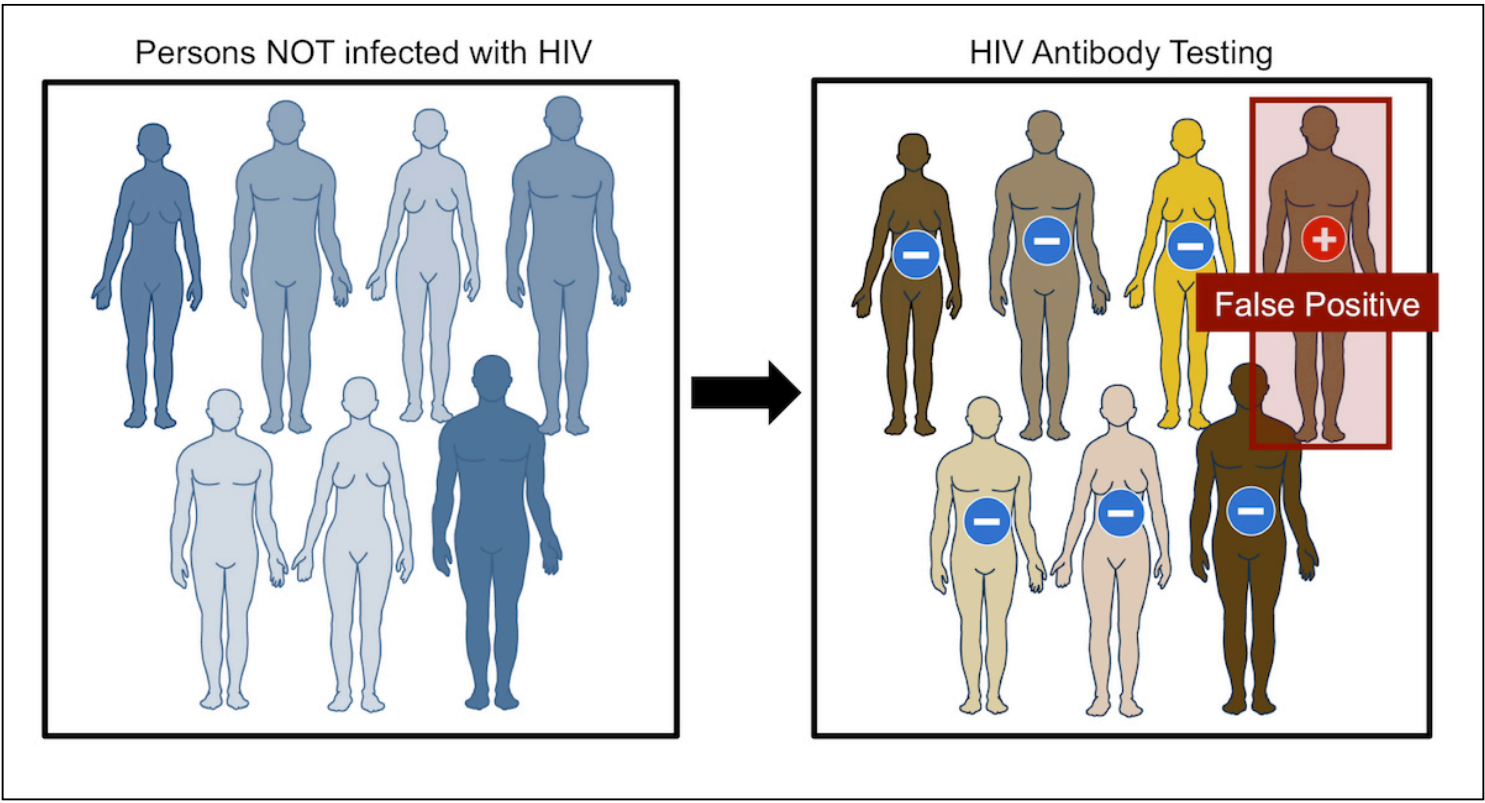


Figure 16 Diagnostic Test Performance in Persons with Acute HIV

During acute HIV (shown in shaded area), the typical pattern is positive HIV RNA, positive HIV p24 antigen, and negative anti-HIV antibodies. Note that with very early acute HIV, the HIV p24 antigen test may be negative. The colored circles indicate when the test typically becomes positive (blue for HIV RNA, green for HIV p24 antigen, and purple for HIV antibody).

Illustration: David H. Spach, MD

